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“Endothelial Progenitor Cells” as a Therapeutic Strategy in Cardiovascular Disease

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Abstract:

Cardiovascular atherosclerotic and ischemic diseases constitute the leading cause of morbidity and mortality throughout middle- and high-income countries. More efficient preventive and regenerative therapeutic strategies are therefore urgently needed. The repeated finding that putative “endothelial progenitor cells” (EPCs) can efficiently promote angiogenesis and restore perfusion of ischemic tissues has provoked a wealth of studies evaluating and developing their therapeutic potential. In the present review, we discuss the growing knowledge about various distinct cell populations which have been collectively termed “EPCs”, including myeloid cells and progenitor cells of different origin. We also present clinical studies aiming to examine their therapeutic potential for cardiovascular disease. In addition, we will discuss recent insights into mechanisms leading to dysfunction of “EPCs” in cardiovascular disease. Those findings may help to optimize autologous cell-based treatment approaches, as well as to establish cellular dysfunction itself as an interesting novel therapeutic target.

Keywords: Cell therapy, cardiovascular disease, cell dysfunction, endothelial progenitor cells.

1. INTRODUCTION

Impaired endothelial cell integrity and function play a major role in atherosclerosis, as well as in ischemic cardiac and peripheral disease. The need to maintain endothelial function and integrity throughout life is challenged by the constant exposure of the endothelium to recurring noxious influences, such as cigarette smoking, dyslipidemia and hyperglycemia that induce increased endothelial reactive oxygen species (ROS) generation ([1, 2], reviewed in [3]).

Cardiovascular pathologies may result from deterioration of endothelial function and integrity in both, the microcirculation as well as the macrocirculation. As microvascular endothelial cells (EC) become dysfunctional and apoptotic e.g. in patients with diabetes mellitus (DM) or hypertension, capillaries in vital organs as well as in limbs dissociate, which results in inadequate tissue perfusion and impaired wound healing [4, 5]. In the macrovasculature, endothelial dysfunction i.e. impaired endothelial nitric oxide (NO) availability, is a hallmark of early atherosclerosis. Today's conception is that constitutive or recurring endothelial activation, in particular in regions of oscillatory shear stress or turbulent blood flow, is characterized by upregulation of adhesion molecules, loosening of EC-EC junctional bonds, increased generation of ROS and impairment of NO availability, which finally induce and maintain an increased inflammatory state of the vascular wall. As a result, the enhanced recruitment of leukocytes and inflow of plasma lipids and lipoproteins into the sub-endothelial space promote the formation of atherosclerotic plaques [6]. Progressing plaque growth, together with plaque rupture or erosion and thrombosis induce acute vessel occlusion, such as in myocardial infarction (MI) or ischemic stroke. While acute treatment focuses on the restoration of tissue perfusion e.g. by thrombolysis or percutaneous intervention, the induction of reparative angiogenesis i.e. the growth of new microvessels into the tissue at need, is considered a promising strategy to confine post-infarction cardiomyocyte (CM) loss. This strategy crucially relies on the endothelium as a main player.

Although today we have several pharmacological tools available to combat the progression of CVD and to enhance survival following acute cardiovascular events, a true regression of atherosclerosis as well as cardiac repair after MI or in ischemic heart disease are not sufficiently achieved by currently established treatments. Therefore, different cell-based therapies i.e. transplantation of immature cell types, such as stem and progenitor cells (S/PC) have been investigated for ischemic organ or tissue repair with great enthusiasm [7]. While the initial high hopes placed in this treatment have not yet been clearly fulfilled, we have gained first clinical experience, mostly with autologous bone marrow cells (BMC), including the establishment of safety of the procedure and the achievement of a significant, though rather small improvement of cardiac function after an acute MI. We have also achieved a better understanding of the cellular and molecular mechanisms underlying the action of various cell populations, including the so-called "endothelial progenitor cells" (EPC). As described below, this term has been used for several different cell populations, and not all of these are "true" endothelial progenitors (in the sense that they differentiate into mature endothelial cells *in vivo*), but may rather promote endothelial growth in a paracrine fashion. The field is now advancing towards an optimization of treatment protocols, which are better adjusted to the individual pathology and characteristics of the donor/recipient, and is focusing on overcoming important limitations of cell-based therapies, including ways to establish *ex vivo* technologies to improve the functional capacity of patient-derived autologous cells.

2. WHAT ARE “ENDOTHELIAL PROGENITOR CELLS”?

In the early 1990s, endothelial cell progenitors were largely examined in embryonic development of the cardiovascular system and the term was applied to cells with a low degree of differentiation, but a high potential to differentiate towards endothelial cells. These cells were also termed angioblasts [8]. In 1997, Asahara and colleagues reported the existence of circulating cells with endothelial lineage potential - i.e. putative “endothelial progenitor cells” – in the postnatal setting i.e. within enriched CD34+ cell fractions obtained from the peripheral blood of human donors [9]. Despite the in vitro endothelial differentiation potential of postnatal CD34+ cells, only few of these cells or their progeny were detected at sites of neovascularisation after transplantation in in vivo ischemia models. In comparison, the observed functional effects (considerably increased neovascularization and blood flow) were beyond what could be expected from differentiated progenitor cells replacing lost EC, or generating new ones [10]. Further findings underlined the importance of paracrine effects of the putative “EPCs” for the observed functional improvement i.e. reduction of ischemia [11-13].

Methodological and terminological inconsistencies have contributed to the fact, that several different cell populations have been termed “endothelial progenitor cells”: After the late nineties, so-called “EPCs” were obtained by subjecting unsorted peripheral blood mononuclear cells (PBMC) to adherence culture in endothelial growth medium immediately, omitting the CD34+ enrichment step [14]. The resulting cells had both, monocyte markers such as CD14, and endothelial markers such as VE-cadherin [14]. Later work verified the myeloid identity of the outgrowing cells [11, 15]. However, the majority of the community adopted the term “endothelial progenitor cells” for these PBMC-derived cells and also discussed their possibilities in this context, fuelled by the cells’ therapeutic capabilities which was by now beginning to be verified in human patients [14, 16, 17].

Minute characterization of PBMC-derived cells has established that two - still heterogeneous – populations of cells with angio-supportive characteristics can be obtained from adult peripheral blood: “early EPCs” or “early outgrowth cells” (EOC), which adhere to fibronectin-coated plastics within 4-7 days, at which time colony-like cell clusters can be observed; and “late EPCs”, also termed “endothelial colony-

forming cells (ECFC)”, which adhere to collagen-coated plastics faster (within two days), but take longer (two to three weeks) to form proliferating colonies [13]. EOC (or “early EPCs”) mainly consist of monocytic cells, which do not give rise to mature EC, but integrate into the subendothelial space of an injured host vessel wall after transplantation - a feature that reminds of resident “patrolling” monocyte subtypes [18, 19]. In contrast, ECFC (or “late” EPCs) contain only minimal amounts of myeloid cells, while a larger portion of them can differentiate into cells of endothelial phenotype [12, 13, 20, 21]. Similar functional characteristics seem to apply to CD34+ vs. CD14+ circulating cell types, freshly isolated from the circulating blood [22]. Of note, the role of monocytes indeed seems to be fundamental to the architecture of newly generated vascular networks, due to their ability to form linear and branched structures in three-dimensional matrix and to interact with various cell types of vascular and non-vascular identities, thus guiding the course of the neovessel [23].

Apart from circulating cells, derived from the bone marrow (BM) or other organ stem cell niches, various types of vascular resident S/PC have been identified. They may contribute to angiogenesis,

population of vascular grafts by recipient cells, and possibly also to vessel wall remodelling and atherosclerosis [24-28]. Their role in physiology and pathophysiology seems to be underestimated as far as now, especially in the context of stent healing or in-stent restenosis [29].

For this review, the term “endothelial progenitor cell” will be applied as a generic term including “true” endothelial progenitors - circulating and resident - which are able to give rise to endothelial cells as well as to myeloid EOC, that rather act in a paracrine fashion on the endothelium. For clinical studies, we will largely refer to experience with autologous BMC, although they clearly constitute a heterogeneous cell population containing mature cells in addition to mesenchymal and hematopoietic progenitors (MSC, HSC, respectively).

This concept reflects the two directions in which research on “EPCs” progresses: (1) the ongoing search for the “true” angioblast/endothelial progenitor cell, driven by the motivation to understand underlying cellular processes of vascular pathologies and regeneration, as well as to identify a defining set of antigenic markers [30], and (2) the clinically motivated search for the therapeutically most efficient cell(s), not necessarily willing to await improved mechanistic understanding, but with the chance to speed up the development of potentially efficient therapies. Yet, mechanistic understanding, including cellular identity, is critical to clinical application, in order to improve efficiency and safety, while economic, ethical and logistic aspects might hinder the transgression of specific cell types into the clinical setting, which are nevertheless highly suited in in vitro and pre-clinical studies, such as embryonic SCs.

3. THERAPEUTIC POTENTIAL AND NON-TARGET EFFECTS OF “ENDOTHELIAL PROGENITOR CELLS”

The contribution of different types of endothelial supportive cells to tissue healing and/or regeneration varies between individual pathological settings. Maintenance or restoration of a confluent, non-activated endothelial layer requires long-acting mechanisms considering also the delicate interactions between various cell types. Monocytes/EOC via their capacity to phagocytose dysfunctional or dying cells, to export cholesterol from the vessel wall, and to secrete distinct spectra of cytokines, play an important role here [11-13, 31, 32]. In contrast, regeneration of the postinfarction myocardium, again crucially involving the phagocytotic and paracrine activity of monocytes, progresses in two distinct stages: first, a fast “clean-up” of dead and dying cells, and later on, the stimulation of repair i.e. angiogenesis and thereby enhancement of tissue perfusion [33, 34]. Other EOC- and BMC-mediated mechanisms include remodelling of the extracellular matrix and possibly the enhancement of cardiomyocyte (CM) survival and functionality by paracrine mechanisms [35-39].

The qualitative and quantitative composition of the “EPC”- or BMC-derived secretome crucially affects short and long-term therapy success. In general, cytokines secreted by the recruited leukocytes are considered to beneficially impact on infarct size and cardiac function after MI [40]. On the other hand, leukocyte-derived cytokines have been associated with post-MI microembolisation and may affect vessel maturation [41-47]. Following MI, the time interval between the acute event and cell application is characterized by alterations in milieu conditions i.e. a high inflammatory load within the first week

followed by a more repair-promoting milieu at later stages [33, 34]. BMC transplantation after the first week post-MI could therefore be more effective [48], a concept which is currently examined prospectively in clinical trials in patients after MI (SWISS-AMI, TIME).

As “EPCs” may support vascularisation of an ischemic tissue, so they may do with a (silent) tumour, potentially facilitating tumour growth [49, 50]. In this context, the risk of de novo tumour formation/leukaemia induction, which has been observed after transplantation of foetal stem cells or cord blood cells [51, 52], might be reduced by using committed progenitor cells, or differentiated (autologous) adult-derived cells [53, 54]. Although angiogenesis in the ischemic heart or peripheral limbs might be a therapy target of BMC/“EPC” transplantation, angiogenesis at developing or mature plaques is unwanted, as it may confer plaque instability [55-57]. Even in the absence of noticeable angiogenesis, recruited HSC/myeloid EOC may facilitate intima thickening, destabilize plaques or promote calcification [58-61]. Likewise, circulating or resident progenitor cells may contribute to intima thickening [62]. Although, in the current clinical trials of BMCs in patients after MI, no safety concerns have been raised, this aspect always needs to be taken into account for cell-based therapies.

Immunological aspects hinder the usage of allogenic donor cells, while embryonic stem cells -although harbouring favourably high differentiation potential and low immunogenic risk - are potentially tumourigenic and raise ethical concerns, further aggravated by the opposition of high numbers of potential recipients and the low number of in-hospital abortions. From ethical, economical and safety aspects, autologous cell sources therefore remain as the most practicable cellular source. Indeed, cells with angiogenic/vascular potential have been detected even in CVD patients, albeit cellular functionality negatively correlates with cardiovascular risk, leaving the least potent cells to the patients who would need the most functional ones [63-65]. Two strategies can be derived from those considerations: the protection of stem cell niches throughout life needs to be addressed prospectively [66]. For the individual patient in need for treatment, ex vivo “priming” or engineering of autologous cells might provide more efficient approaches in the near future. Various strategies have been devised towards that end, including improvement of isolation and storage protocols for the selection of the most efficient cell type and their functional preservation [67, 68] as well as ex vivo pharmaceutical pre-treatment or genetic engineering of cells prior to re-implantation. Improving the number of cells to reach the target tissue and to persist there e.g. by optimizing the route of transfusion or accompanying pharmacological treatment (e.g. CD26 inhibitors), might further improve efficiency. Below, we will discuss distinct strategies to employ endothelial growth promoting cells in distinct pathological situations.

3.1. “EPCs” to Treat Microvascular Dysfunction and Rarefaction

Microvascular rarefaction i.e. the reduction and loss of a functional network of capillaries normally ensuring organ perfusion, likely contributes to different forms of cardiomyopathy, including ischemic or hypertensive cardiomyopathy and diabetic cardiomyopathy. Deteriorating microvasculature loses pericyte coverage and assumes a tortuous phenotype with discontinuous endothelium (Fig. 1) [69]. Although at the first glance paradoxical, the loss of capillaries in heart, kidney and peripheral tissues shares some molecular and cellular mechanisms with diabetic retinopathy, such as increased capillary permeability and tortuosity (reviewed in [70]).

The demonstration of the pro-angiogenic potential of “EPCs” has therefore raised hopes for a novel treatment strategy, especially in those patients who do not respond to other modes of therapy [16]. However, cellular functions of “EPCs” are severely affected i.e. in diabetic patients and smokers, thus potentially limiting the efficiency of autologous cell transplantation [64, 65]. Ex vivo modification of cellular function prior to re-introduction (Table 1) might therefore harbour specific benefits for patients with an impaired “EPC” function. As an example, the overexpression of platelet-derived growth factor (PDGF), a mediator of pericyte-EC interaction during microvessel stabilization, has been proposed. Interestingly, ex vivo PDGF treatment of human CD34+ progenitor cells enhanced their homing in a diabetic mouse model and promoted in vitro the emergence of adherent cells/clusters that expressed the endothelial markers VE-cadherin and CD31 [71]. Ex vivo overexpression of growth factors (e.g. vascular endothelial growth factor (VEGF), insulin-like growth factor (IGF-1)) [72, 73], oxidative defence effectors (e.g. manganese superoxide dismutase), endothelial nitric oxide synthase (eNOS), or PPAR α activators served to improve cellular functions in in vitro studies and animal models [72-76]. Transfer of these experimental approaches into the clinical setting might come closer with the development of safer vectors and the availability of first results from clinical trials employing direct and cell-mediated gene therapy (reviewed in [77, 78]). Alternatively, biodegradable matrices could be adopted as a means for time and dosage-dependent release of mature proteins e.g. for topical application in diabetic wound healing [79]. Furthermore, systemic effects are exerted by lifestyle approaches, such as physical exercise training or flavonoids in certain foods (tea, red wine, chocolate), as well as drugs with pleiotropic actions. Especially statins have been shown to be effective in improving “EPC” availability and function via activation of the PI3K/Akt/eNOS pathway, besides potent anti-inflammatory effects and blockade of the rate-limiting enzyme of cholesterol synthesis, HMG-CoA reductase [80-82].

The kallikrein-kinin-system has gained attention for its role in regulating angiogenesis, as well as vascular stabilization [83-85]. Cellular players are mature cells of the vasculature, such as EC or SMC, but also EOC, as well as individual progenitor and leukocyte subtypes, which regulate angiogenesis in a paracrine manner. In a non-diabetic mouse model, expression of the kinin B2 receptor was necessary for the recruitment of progenitor cells to ischemic tissue and initiation of capillary growth and reperfusion [86]. In vitro, EOCs migrated towards the B2 receptor ligand bradykinin (BK) and were enriched out of the heterogeneous PBMC by using BK as a chemoattractant. Progenitor cells from patients with CVD could not be enriched by BK-induced migration [86]. Interestingly, progenitor cells and monocytes from young type 1 diabetic patients without CVD retained B2 receptor expression and BK-induced migration, while individual B2 receptor-mediated cellular functions, such as NO generation, were impaired [87]. Uncoupling of the B2 receptor from downstream signaling was associated with reduced expression of tissue kallikrein, a protease mediating kinin generation and matrix degradation, leading to impaired invasive capacity of EOCs from type 2 diabetic patients [88]. Consequently, ex vivo overexpression of both, kallikrein and the B2 receptor, in diabetic EOC did restore invasive capacity [88]. While interactions between the members of the kallikrein-kinin system may be complex and simple overexpression of one component might not suffice to restore angiogenesis in certain pathologies [88, 89], combinatory treatment, together with re-coupling of eNOS, might enhance EOC pro-angiogenic capacity.

Maturation of the nascent vascular network i.e. the recruitment of pericytes and SMC to provide stability and reduce permeability is crucial to ensure functionality of newly generated vasculature [90]. To achieve both, initiation of angiogenesis and subsequent pericyte recruitment, VEGF and PDGF have been successfully applied as dual gene therapy in various animal models [91, 92]. Recent work furthermore indicates an additional role of the kinin B2 receptor in vessel maturation by pericyte/SMC recruitment [84]. The intrinsic capacity of “EPCs”/EOCs to counteract capillary permeability by stabilizing tight junctions, as was shown recently for myeloid multipotent progenitor cells, might be enhanced by tailored modification of autologous “EPCs” paracrine activity prior to reintroduction e.g. by cotransfection with different growth factor genes [93].

Besides 'classical' paracrine substances, further mediators of endothelial function, supplied by “EPCs”, as well as alternative routes of delivery have been identified. Interesting novel data have indicated that actively shed membranous vesicles, depending on their size classified into microparticles (MP), exosomes, or microvesicles, may carry soluble proteins, membrane proteins, lipids and nucleic acids, thus mediating communication between the cell of origin and the target cell over variable distances [94-98]. Exogenously modified “EPC”, which shed MPs, or the “EPC”-derived MPs themselves are therefore considered promising tools for targeted drug delivery [99]. This principle could be extended to ex vivo engineering of “mother cells” shedding MPs containing a desired protein or microRNA profile. The cells would then be available for producing multiple doses of paracrinally active “conditioned medium” to be transplanted during several treatment sessions, which might increase efficiency of the therapy [100].

Therefore, by their intrinsic paracrine activity, “healthy” EOCs might counteract the destabilization of microvascular networks, as well as support the re-constitution of a mature, stable network of new capillaries (Fig. 1). Ex vivo engineering or pre-conditioning of patients autologous EOCs (i.e. by overexpression of pro-survival factors and/or chemoattractants, or exposure to survival-promoting drugs (Table 1)) prior to transplantation might be helpful, not only to promote angiogenesis, but also to enhance stability of newly generated or pre-existing micro-vessels.

3.2. Potential of “EPCs” as a Therapeutic Strategy for Macrovascular Disease

As for the establishment of a functional neovasculature, the interplay between endothelial and sub-endothelial cells of the vascular wall is critical for the maintenance of macrovascular function. During life, various factors, such as circulating lipids and glucose, turbulent flow due to vascular geometry, and inflammatory cells activated by environmental toxins, add up to establish permanently inflamed areas of the vascular wall, predominantly at bifurcations. This is followed by intima thickening due to the immigration of leukocytes from the blood and SMC from deeper layers of the vessel wall. Inflammation is further promoted by leukocytes entering the plaque via vasa vasorum. The loss of endothelial function and integrity plays a crucial role in atherogenesis: Endothelial-derived NO generation, important for controlling SMC relaxation and proliferation, is reduced and the tight coupling of endothelial cells through junction molecules, as well as low expression of adhesion molecules, are reverted. In consequence, plasma proteins can leak into the subendothelial space, followed by leukocytes which invade the vessel wall. Therefore, the reconstitution of endothelial confluence and quiescence is regarded key in prevention and halting of atherosclerosis, while its reversal might be achieved by enhancing reverse cholesterol transport e.g. by HDL [101]. Towards this end, functionality

and survival of EOC when exposed to the pro-apoptotic conditions within the inflamed vessel wall are critical for effectively counteracting atherosclerosis [102].

The role of “EPCs” in counteracting, but also potentially in promoting atherosclerosis seems to be conveyed mainly by their paracrine action Fig. (2). They can attenuate vascular inflammation e.g. by interleukin-10 secretion, support endothelial cell turnover e.g. via NO generation or secretion of interleukin-8, hepatocyte growth factor or VEGF [11], or by promoting reverse cholesterol transport [103]. Special attention has been given to the capacity of EOCs for NO generation, which is considered a hallmark of their functional capacity [19, 104-106]. Indeed, “EPC” may lose the capacity for NO generation well before the onset of vascular disease in persons with increased risk, such as diabetes mellitus or pre-hypertension [19, 87]. Paracrine dysfunction of “EPC” in persons with enhanced cardiovascular risk (e.g. in smokers or diabetic patients) therefore may impair endothelial functions two-fold [107-109]. In the chronic setting, treatment options such as statins or exercise training, which can improve “EPC” paracrine actions in vivo without need of cell extraction, ex vivo modification and re-implantation, seem favourable, as drugs/lifestyle modifications can be applied over long time courses. In contrast, single cell transplantation treatments may only cover shorter time periods. However, cell-derived non-immunogenic products, such as microvesicles loaded with specific proteins or microRNA, might be suitable for repeated administration. Detailed investigation of microvesicle composition and the systemic effects exerted by individual microRNAs in CVD are therefore awaited with great expectations.

As discussed elsewhere [110], percutaneous coronary intervention (PCI) induces severe damage to the endothelium i.e. endothelial denudation and stretching of matrix and underlying cell layers. The fast re-establishment of a confluent, quiescent endothelial layer is therefore accounted crucial for both, the attenuation of atherosclerosis, as well as the prevention of in-stent re-stenosis/thrombosis. Antibody-mediated capturing of circulating CD34+ progenitor cells by implanted stents is currently followed as a potential strategy to promote endothelial healing after stent implantation [111, Current Vascular Pharmacology, 2012, Vol. 10, No. 1 113

112]. Although safety and feasibility were suggested by several studies [113-115], higher rates of target vessel failure in patients receiving the “EPC-capture-only” stent were described for high risk lesions as compared to drug-eluting stents [116-121]. Therefore, a new generation of combined CD34+ attracting and drug-eluting stents is currently entering clinical trials (REMEDY, REMEDY-OCT) with the hope to achieve accelerated endothelial healing without increasing restenosis risk [122].

While progenitor cells circulate in the blood only in low numbers, resident S/PC within adjacent vascular sections provide a more readily available source for re-population of vascular grafts, and possibly also for the reendothelialisation of stents or - with adverse result - intima growth [24, 25, 27, 123, 124, 28]. Vascular resident S/PCs are positioned where they can rapidly provide support to damaged endothelium, but naturally they will be equally affected by blood-borne stressors, like inflammatory mediators increasing oxidative stress within the vessel wall, and may therefore suffer equally [125]. Consequently, bone marrow-derived progenitor cells, leaving their protective environment and entering the circulation, as well as vascular-resident progenitor cells, will both be exposed to different factors, leading to cellular dysfunction [126].

Statins achieve reduction of cholesterol levels by blocking HMG-CoA reductase, but they also activate the pro-survival Akt/eNOS pathway and skew EOC differentiation towards a repair-promoting phenotype [81, 127]. In the setting of stent implantation, concomitant statin therapy serves to accelerate re-endothelialisation and reduce intima thickening and thrombosis, improving survival after PCI [128-131]. Contributing to those effects might be the increased mobilization, as well as better homing characteristics of EOC and progenitor cells (Table 2) [127, 132, 133].

Similarly, physical exercise training improves endothelial function in various physiological and pathological settings [134-136]. Exercise-induced effects are mainly ascribed to a systemic decrease of inflammatory agents, increased NO availability, downregulation of NADPH oxidase and upregulation of enzymes involved in antioxidative defence, but recent studies have also established higher numbers of circulating "EPC" [137-145]. Aortic valve stenosis represents another major cardiovascular pathology, which shares underlying mechanisms with atherosclerosis, such as leukocyte infiltration and endothelial deterioration, but calcification processes play a larger role than in atherosclerosis [146, 147]. The progression of valve disease, characterized by EC loss, coincides with reduced survival and lower circulating numbers of "EPC" [148]. In this context, exercise training served to prevent aortic valve disease in a high-cholesterol induced mouse model of aortic valve disease, albeit only associations exist about the role of increased circulating "EPC" levels in the preservation of valve integrity [149]. In contrast to "EPC", circulating cells which co-express osteocalcin and bone alkaline phosphatase, were identified to actively promote calcification hyperglycaemia and diabetes. Of note, those myeloid calcifying cells (MCC) were distinct from HSC/"EPC" according to their antigenic profile, including lack of CD34 expression [150]. However, the derivation of MCC from cultured PBMC, albeit under growth conditions distinct from EOC, suggests inclusion of MCC screening into the panel of characterization for EOC used for cell therapy. Lack of comparable characterization data currently precludes conclusions about the modulation of MCC in CVD and therapy strategies, such as statin treatment or exercise training [150, 151]. Resident vascular cells, such as SMC or pericytes might also differentiate into calcifying cells under certain milieu conditions [152]. Exercise-mediated maintenance or restoration of endothelial confluence - possibly via "EPC"-mediated mechanisms - might help to counteract this process [151].

Much attention has been paid to the role of diet in retaining endothelial integrity and function, including unsaturated fatty acids, carbohydrate content and flavonoids [153-155]. Flavonoids present in red wine or cocoa, such as resveratrol or epicatechin, increase NO generation and reduce oxidative stress [156-158]. In vitro, as well as in pre-clinical and clinical studies, red wine, but not ethanol, enhanced "EPC" survival and increased vascularisation of ischemic tissues, mostly ascribed to NO mediated mechanisms [156, 158-160]. In human CAD patients, intake of flavanol-rich cocoa increased NO production and the abundance of circulating "EPC" and improved endothelial function [161].

In summary, EOCs as well as circulating and vascular-resident S/PC may protect from atherosclerosis by preserving critical endothelial functions. Due to the chronic course of plaque development, the beneficial effect of single cell application might turn out to be limited. Furthermore, the availability of pro-atherosclerotic and calcifying elements within the heterogeneous populations of PBMC and BMC, bears the risk of advancing, rather than counteracting atherosclerosis. Therefore, modulating the systemic condition towards a reduction of inflammation and strengthening of antioxidative defence

and promoting reverse cholesterol transport via systemic treatments holds greater promise. Resident or circulating “EPCs” will be among the cells benefiting, with niches protected, trafficking improved and differentiation driven away from pro-inflammatory directions.

3.3. “EPCs” for Cardiac Repair

The preservation of functional myocardium after AMI requires a fast restoration of reperfusion, as well as supplementation of contractile and structural elements i.e. cardiomyocytes (CM) and fibrocytes. Due to their ability to enhance angiogenesis in ischemic situations, EOC, as well as various other BM-derived (stem) cell populations are regarded as promising tools for post-MI therapy [9, 162-164].

While in individual clinical studies and meta-analyses a small, but significant benefit of cell therapy for left ventricular (LV) function has been suggested, not all trials were able to detect significant improvement in cardiac function [165-171]. The differential outcome of those studies has been attributed to technical differences, especially isolation and transport/storage procedures, yielding cell populations with distinct composition and functional capacity, but also to the time point and mode of application, as well as patients characteristics [48, 67, 172].

For practical reasons, most clinical studies have employed total BMC, whose cellular composition is rather mixed, including e.g. MSC, HSC, and stromal components. In consequence, various cellular and molecular mechanisms

- in detail explored by using more defined cell populations in vitro or in animal models - might contribute to the observed salvage on myocardial function in patients receiving autologous BMC. “EPC” differentiation into functional CM is still debated and the main effect of BMCs in myocardial salvage after MI is likely held by promotion of angiogenesis [162, 173-184]. For MSCs contained within the BMC some cardiopoietic potential, as well as the capacity to affect conductance, have been suggested and appears to be promoted by a “cocktail” of cytokines [185, 186]. Few comparative clinical studies have investigated the relative potential of total BMC in comparison to more defined cell populations, with heterogeneous outcome. Similar efficiency was attested to EOC or CD34+CXCR4+ cells as compared to unselected BMC [164, 167, 187, 188]. To overcome low engraftment and in order to deliver a relevant number of efficient cells, high numbers of total BMC are usually transplanted. Thereby, higher absolute numbers of contaminating, potentially inflammatory cells might be coinjected, possibly impeding the healing effect and triggering adverse events [188, 189]. To further optimize the efficacy of cell engraftment after transplantation, various routes of cell delivery have been used, including intracoronary, direct intracardiac (surgical transepicardial or catheter-based transendocardial) or intravenous injection. The appropriate method likely depends on the clinical scenario (i.e. acute infarction vs. chronic ischemic disease) and on the cell types used. Some studies have suggested a higher cell retention rate after intramyocardial injection, which allows targetted delivery, aided by mapping systems [190]. However, cell retention is generally low: Even with direct intramyocardial injection, only around 10% of the injected cells are retained within the first hour of transplantation, with a further decline over the next days [191, 192]. Several groups therefore currently focus on enhancing the retention of injected cells, including delivery of cells within matrices, improvement of the microenvironment (i.e. recapitulating “niche-like” conditions), and ex vivo cell modifications (e.g. induction of adhesion molecules, improvement of survival) [193-199].

Recent meta-analyses suggest that transplantation of total autologous BMCs may improve LVEF, but not left ventricular remodelling, and their potency depends on the donor/recipient characteristics, with active smoking, male gender, age and diabetes negatively influencing efficiency [200-203]. As donor and recipient necessarily are the same person, it is difficult in clinical studies to discern the impact of the milieu, the cells are exposed to after implantation, from intrinsic dysfunctions of the cells. In vitro studies and ex vivo analysis of transplanted cells indicate that both are relevant. Transplanted cells and their surroundings mutually interact, and therefore improving transplanted cells' function ex vivo (Table 1) together with pharmacological and/or lifestyle-based lowering of systemic inflammatory status (Table 2) might be necessary to increase efficiency of autologous cell-based therapy after myocardial infarction.

Myocardial remodelling after acute infarction is characterised by two distinct phases in short to mid-term, with fibrotic processes stretching on to longer time periods [33, 204]. The mainly inflammatory conditions within the first week, characterised by apoptosis/necrosis and phagocytosis, in opposition to regenerative/angiogenic processes which occur during the following weeks, might serve to explain why cell application may be more effective in improving LVEF at 6 days as compared to 3 days after hospitalization [48]. After establishment of cardiomyopathy, injection of autologous BMC was still able to reduce infarct size and improve ventricular function [205]. Similar to other cell types, EOC or progenitors become dysfunctional and apoptotic when exposed to high levels of TNF- α or IL-1 α , conditions occurring immediately after aMI [206, 207]. Transplanted cells further suffer from the still ischemic conditions at the target site [208]. In contrast, inflammatory cytokines decrease in the second and third week after infarction, supporting "EPC" persistence. In the blood of patients successfully undergoing cardiac rehabilitation, increased availability of CD34⁺KDR⁺ PC was described, which correlated with systemic VEGF, IL-8 and IL-10 levels [209]. Therefore, conditions early after infarction appear to be detrimental to cell survival and function, explaining better effects with later time points of transplantation [48]. Vice versa, dysfunctional PC/EOC may also supply inflammatory cytokines, with adverse effects on myocardial healing and possible arrhythmogenic potential through paracrine stimulation of adjacent CM [185, 210]. Therefore, combination of cell application with statin therapy might yield synergistic effects, due to the systemic anti-inflammatory effects of statins, as well as specific effects on "EPC" survival and function e.g. differentiation [81, 206, 211]. Physical exercise, through multiple cellular and molecular mechanisms, including antioxidant and anti-inflammatory effects, is able to improve measures of cardiac function and geometry in heart failure patients and might also increase the success of stem cell transplantation [212]. In addition to their paracrine effects, electrophysiological alterations after cell transplantation have been debated and this depends on the cell type used [213]. Mechanistically, a lack of electrical coupling and distinct conductance characteristics between injected cells and the surrounding myocardium has been suggested to underlie arrhythmias, as suggested by various preclinical models i.e. co-culture and animal studies [214, 215]. Both, the lineage identity and stage of differentiation of the transplanted cell [216-218], as well as the position it is deposited at (surrounding myocardium viable or dead? [219]) affect electrical coupling to the host myocardium. For cardiopoietic cell types, the differentiation into various distinct CM lineages with individual action potential furthermore accounts for the occurrence of arrhythmias [220]. However, the electrophysiological implications of cell therapy, as well as underlying cell-type specific

mechanisms are not entirely clear today and several recent clinical studies observed either no signs for pro-arrhythmic effects or even report anti-arrhythmic effects of stem-cell based approaches [221-224].

Due to their paracrine action, EOCs or other progenitor cell types, may affect all cell types in the vicinity, EC as well as CM, local SC niches or fibrocytes. In this context, their effect seems to be limited by the distance to the targeted CMs as well as by the length of their persistence [37, 225, 226]. Hence, higher numbers of recruited/persistent cells and uniform distribution along the target zone are aimed for in the clinical situation [227-231]. To ensure uniform high therapeutic quality of clinical stem cell therapy, optimized and standardized cell preparation, transport, and storage protocols need to be established. First experience on logistics (including equipment and training of personnel) and quality issues are available now from multicentre trials [232]. Finally, relevant outcome parameters defining the success of stem cell therapy and - potentially - its superiority vs. other therapies need to be agreed upon in order to compare individual approaches [233].

Cell therapy after MI, even after optimization of cell function, engraftment and survival, might still meet its limit through the impotency of this therapy approach to supply sufficiently high numbers of structural and contractile elements. Surgical approaches aiming specifically at improving structural stability and contractile force -especially for the long-term prevention of heart failure -include ex vivo engineered patches of various biomaterials, optionally seeded with cells and/or growth factors [234, 235]. Although still facing many difficulties on its way to clinical applications, this approach might prove more efficient in patients with advanced heart failure and a severe loss of contractile cardiomyocytes [236, 237]. The demands upon the material serving as matrix are high: it should not elicit host inflammatory response, feature conductance and mechanical properties similar to the host myocardium, and be stable over long time periods under repetitive physical strain [238]. Therefore, scaffold-free cell patches might constitute an alternative approach [239, 240]. Yet, long-term engraftment and stability issues, as well as connection to blood supply may also complicate the development of scaffold-free cardiac patches, so far limiting patch thickness/size [241]. Another obstacle to overcome for both approaches consists in ensuring proper electrical coupling of the transplanted patches to avoid arrhythmia, similar as discussed above for the transplantation of single-cell suspensions [242]. Here, the maturity of the cell type, determining distinct calcium current profiles, is critical [216-218]. While immature cells over time integrate with their neighbour cells, the risk of tumour formation or calcification is increased when embryonic undifferentiated SCs are used [53, 243]. In contrast, electrical coupling remains problematic with pre-differentiated cardiomyocytes. It remains to be seen whether the ex vivo de-differentiated inducible pluripotent stem (iPS) cells offer a better source for the generation of autologous tissue patches [244-246]. Yet, reprogramming of cells derived from CVD patients might be limited by epigenetic changes and DNA damage due to their levels of oxidative stress. Recent pioneering studies have established that epigenetic modifications are carried over to the iPS cell and that reprogramming may introduce aberrant DNA methylation, potentially affecting functionality and/or genetic stability of the reprogrammed cells [247, 248].

4. SUMMARY AND CONCLUSION

Current clinical data support a small but significant benefit of autologous cell transplantation with respect to cardiac function in the setting of acute myocardial infarction. However, the study outcome was divergent with regard to the extent of benefit, and in some cases did not support superiority of stem cell transplantation. Critical points to optimize in future trials are the functionality of the transplanted cells, milieu conditions within the recipient tissue, and rate of cell engraftment. In the setting of post-MI treatment, the main effect of HSC/"EPC" among the BMCs seems to consist in accelerating angiogenesis, thus enhancing perfusion and providing better survival conditions to remaining cardiac cells. Myeloid cells among the "EPC"/BMC might also modulate milieu conditions, such as the balance between inflammatory and repair-promoting cytokines, or nitric oxide and oxygen radical levels.

Preservation of S/PC niches within the bone marrow and possibly also the modulation of their differentiation, function and survival can be targeted by drugs, such as statins, as well as lifestyle adaptation. In patients with established CVD e.g. statin treatment or physical exercise, was able to increase "EPC" availability within the blood and their cellular functions. Furthermore, CD34+ cell-capturing stents combined with drug-eluting design are currently entering clinical trials, and may represent a novel approach to accelerate stent healing in the coronary artery.

The safety of different cell-based treatment approaches needs to be closely monitored with respect to the occurrence of adverse events, such as arrhythmias or cell transplantation-induced promotion of neointima formation.

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Figures

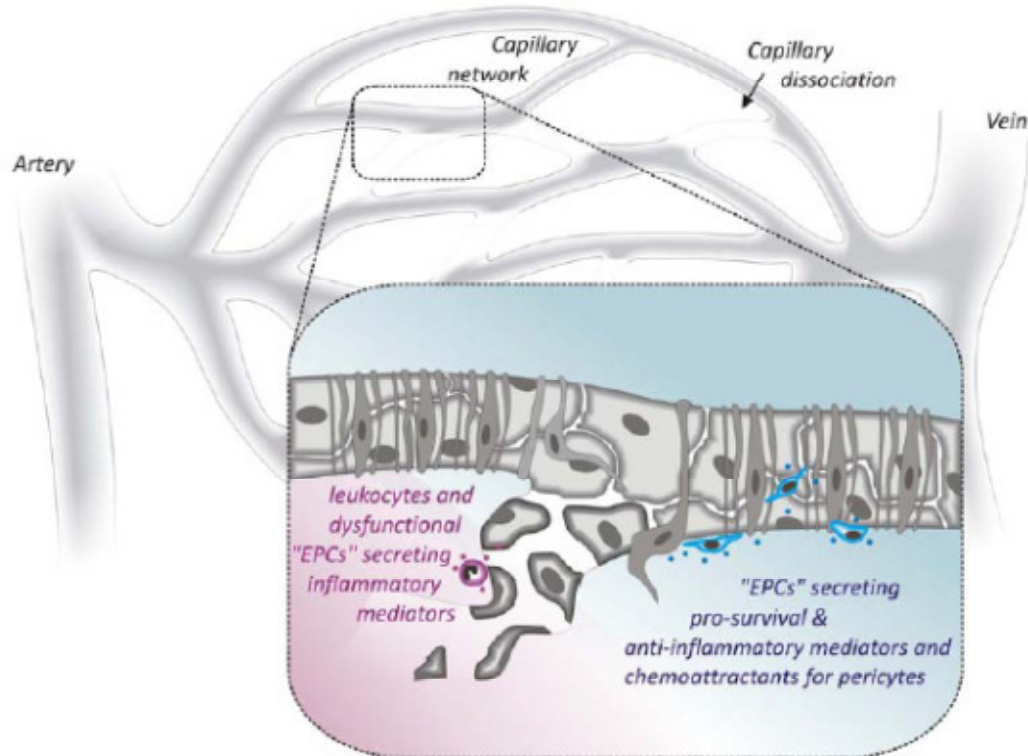


Figure 1. Potential effects of “endothelial progenitor cells (EPC)” on microvascular disease: In conditions with high oxidative stress e.g. in diabetes or aging, microvascular cells (i.e. EC, pericytes) die resulting in dissociation of microvessels. Extravasating leukocytes and dysfunctional “EPC”/early outgrowth cells (EOC) might aggravate this process by the secretion of inflammatory mediators, such as cytokines or reactive oxygen species (purple). Healthy “EPC”/EOC might counteract the loss of endothelial cells in several ways, including secretion of pro-survival factors such as nitric oxide or interleukin 10 and by supporting vessel stability by attracting pericytes (blue).

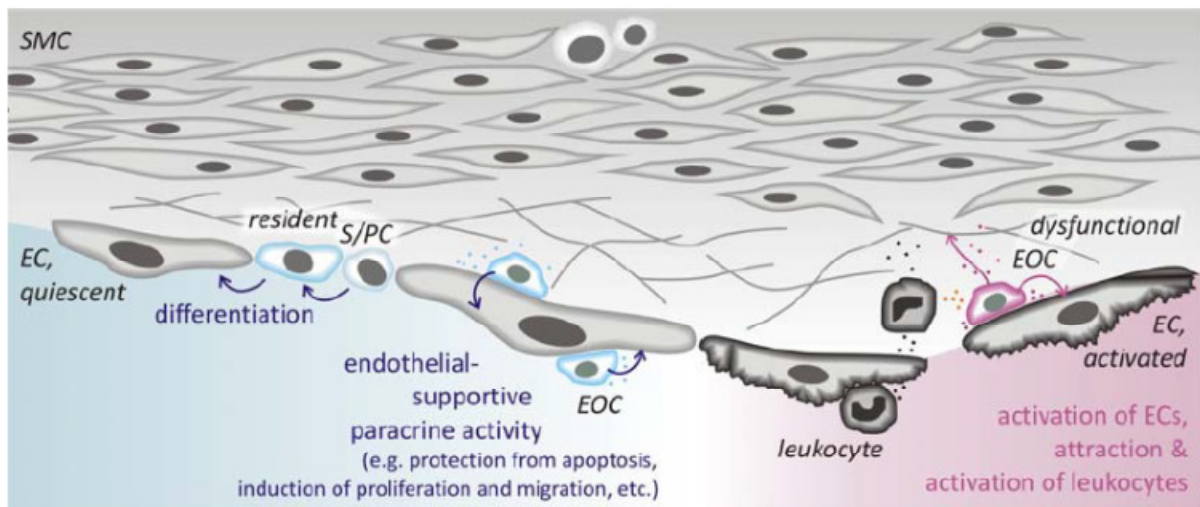


Figure 2: Potential effects of “endothelial progenitor cells (EPC)” in macrovascular disease: Functional “EPC”/early outgrowth cells (EOC) from healthy individuals (blue) are involved in maintaining endothelial integrity e.g. by paracrinally inducing nitric oxide generation or protecting endothelial cells (EC) from apoptosis, while dysfunctional EOC, as e.g. in diabetic patients, might also promote inflammatory responses via secretion of inflammatory mediators and chemoattractants (purple). SMC: smooth muscle cells.

Tables

Targeted Mechanism	Targeted Pathway/Molecule	Outcome
"EPC" recruitment to target tissue (adhesion/invasion)	Growth factors (GF) (<i>ex vivo</i> overexpression in EOC or <i>ex vivo</i> treatment with GF protein, e.g. insulin-like GF 1 (IGF-1) or stromal cell-derived factor 1 (SDF-1))	IFG-1: reduced cardiomyocyte (CM) apoptosis, increased CM proliferation [73] improved migratory capacity, incorporation into forming vascular networks [249] SDF-1: enhanced recruitment of "EPC" [250] enhanced migration [251], improved engraftment/persistence [234] vascular endothelial GF (VEGF): enhanced recruitment to ischemic tissue in mouse model [72]
	PI3K/Akt/nitric oxide (NO) (<i>ex vivo</i> statin treatment, <i>ex vivo</i> overexpression of endothelial NO synthase (eNOS) in EOC)	<i>Statins</i> : enhanced migration [82] reverted loss of EOC adhesive and migratory capacity induced by oxidized low-density lipoproteins (oxLDL) [252] eNOS: increased migration and incorporation into network structures in matrigel [74]
	Hypoxia-induced signalling (<i>ex vivo</i> hypoxia-inducible factor 1a (HIF-1a) overexpression in EOC)	Improved "EPC" homing [253]
	Kallikrein-Kinin-System (<i>ex vivo</i> combined overexpression of human tissue kallikrein and kinin B2 receptor in EOC from type 2 diabetic patients)	Increased invasion of extracellular matrix gel [88]
	Prostanoids (<i>ex vivo</i> treatment of EOC with prostaglandin analogues)	Improved migration [254]
	Cyclooxygenase (COX) (<i>ex vivo</i> inhibition by aspirin, ibuprofen, parecoxib)	COX-1 inhibition either dose-dependently promotes "EPC" migration and adhesion [133] or decreased migratory and adhesive capacity of "EPC" [255]
	Polysaccharides (<i>ex vivo</i> exposure to fucoidan)	Enhanced expression of CXC receptor 4 (CXCR4) on CD34+ cells [256], enhanced migration and network formation [257]
	Growth hormones (<i>ex vivo</i> erythropoietin (Epo) treatment)	Enhanced adhesion to fibronectin [258]
"EPC" survival/ proliferation	angiogenic GF (<i>ex vivo</i> overexpression in EOC or <i>ex vivo</i> treatment with GF protein)	IFG-1: reduced cardiomyocyte (CM) apoptosis, increased CM proliferation, improved cardiac function [73] attenuated cellular senescence [249] VEGF: prevented "EPC" apoptosis [259] SDF-1: attenuates EOC apoptosis <i>ex vivo</i> [260] reduces senescence, increases proliferation [261] platelet-derived GF (PDGF): enhances <i>ex vivo</i> expansion of CD34+ progenitor cells (PC) [71]
	PI3K/Akt/NO (<i>ex vivo</i> overexpression in EOC)	statins: enhanced survival/cell cycling [80] prevented EOC apoptosis induced by oxLDL [252] eNOS: increased ³ H-thymidine incorporation (i.e. proliferation/DNA repair) [74]
	Growth hormones (<i>ex vivo</i> estrogen or Epo treatment)	Estrogen: reduced cellular senescence [262] Epo: enhanced proliferation
	Glycogen synthase kinase 3b (GSK-3b) (<i>ex vivo</i> GSK-3b inhibitor treatment of EOC)	Reduced apoptosis [263]
	Prostanoids (<i>ex vivo</i> treatment of EOC with prostaglandin analogues)	alprostadiol reduced apoptosis, improved proliferation [254]
	Anti-oxidant defense (<i>ex vivo</i> melatonin exposure)	Melatonin induces an overexpression of catalase and superoxide dismutase-1 and improves survival [264]
	Polysaccharides (<i>ex vivo</i> exposure to fucoidan)	Enhanced cell proliferation and migration [257]

Targeted Mechanism	Targeted Pathway/Molecule	Outcome
"EPC" differentiation/function	PI3K/Akt/NO (<i>ex vivo</i> statin treatment, <i>ex vivo</i> eNOS overexpression in EOC, <i>ex vivo</i> eNOS activation by AVE9488)	statins: anti-inflammatory polarization [81] reverted loss of EOC adhesive and migratory capacity induced by oxLDL [252] eNOS: reduction of neointimal hyperplasia [107], enhanced pro-angiogenic activity in mouse model [265]
	Growth factors (<i>ex vivo</i> overexpression in EOC or <i>ex vivo</i> treatment with GF protein)	IFG-1: improved cardiac function [73] SDF-1: better angiogenesis through enhanced recruitment of "EPC" [250] improved angiogenesis induction and preservation of cardiac function [234]
	Hypoxia-induced signalling (<i>ex vivo</i> HIF-1 α overexpression in EOC)	Improved "EPC" homing, enhanced angiogenesis, reduced limb/toe necrosis [253]
	GSK-3 β (<i>ex vivo</i> GSK-3 β inhibitor treatment of EOC)	Attenuated neointima formation and enhanced re-endothelialization after arterial injury [263]
	Cyclooxygenase (<i>ex vivo</i> inhibition)	COX-1 inhibition decreased <i>in vitro</i> pro-vasculogenic capacity of "EPC" [255]
	Prostanoids (<i>ex vivo</i> treatment of EOC with prostaglandin analogues)	alprostadil improved revascularisation [254]
	anti-oxidant defence (<i>Ex vivo</i> manganese superoxide dismutase (Mn-SOD) overexpression, <i>ex vivo</i> melatonin exposure)	Mn-SOD: wound capillarization and closure improved [75], induction of angiogenesis and paracrine action improved [266] melatonin: enhanced network formation of ECs in matrigel in a paracrine manner [264]
	Polysaccharides (<i>ex vivo</i> exposure to fucoidan)	Enhanced cell proliferation, migration and network formation [257]
	Reverse cholesterol transport (<i>ex vivo</i> overexpression of Apolipoprotein A1 in EOC)	Reduction in atherosclerotic lesion area [103]

Table 1: Strategies for Ex Vivo Improvement of Autologous "Endothelial Progenitor Cells" (EPC) for Transplantation. Pre-Clinical And Clinical Results

REFERENCES

- [1] Tyldum GA, Schjerve IE, Tjonna AE, et al. Endothelial dysfunction induced by post-prandial lipemia: complete protection afforded by high-intensity aerobic interval exercise. *J Am Coll Cardiol* 2009; 53: 200-6.
- [2] Vogel RA, Corretti MC, Plotnick GD. Effect of a single high-fat meal on endothelial function in healthy subjects. *Am J Cardiol* 1997; 79: 350-4.
- [3] Cai H, Harrison DG. Endothelial dysfunction in cardiovascular diseases: the role of oxidant stress. *Circ Res* 2000; 87: 840-4.
- [4] Martin JW, Mousa SS, Shaker O, Mousa SA. The multiple faces of nicotine and its implications in tissue and wound repair. *Exp Dermatol* 2009; 18: 497-505.
- [5] Granger DN, Rodrigues SF, Yildirim A, Senchenkova EY. Microvascular responses to cardiovascular risk factors. *Microcirculation* 2010; 17: 192-205.
- [6] Libby P, Ridker PM, Hansson GK. Inflammation in atherosclerosis: from pathophysiology to practice. *J Am Coll Cardiol* 2009; 54: 2129-38.
- [7] Strauer BE, Brehm M, Zeus T, et al. Repair of infarcted myocardium by autologous intracoronary mononuclear bone marrow cell transplantation in humans. *Circulation* 2002; 106:1913-8.
- [8] Risau W. Differentiation of endothelium. *FASEB J* 1995; 9: 926-33.
- [9] Asahara T, Murohara T, Sullivan A, et al. Isolation of putative progenitor endothelial cells for angiogenesis. *Science* 1997; 275:964-7.
- [10] Schatteman GC, Dunnwald M, Jiao C. Biology of bone marrow derived endothelial cell precursors. *Am J Physiol Heart Circ Physiol* 2007; 292: H1-18.
- [11] Rehman J, Li J, Orschell CM, March KL. Peripheral blood "endothelial progenitor cells" are derived from monocyte/macrophages and secrete angiogenic growth factors. *Circulation* 2003; 107: 1164-9.
- [12] Sieveking DP, Buckle A, Celermajer DS, Ng MK. Strikingly different angiogenic properties of endothelial progenitor cell subpopulations: insights from a novel human angiogenesis assay. *J Am Coll Cardiol* 2008; 51: 660-8.
- [13] Yoder MC, Mead LE, Prater D, et al. Redefining endothelial progenitor cells via clonal analysis and hematopoietic stem/progenitor cell principals. *Blood* 2007; 109: 1801-9.
- [14] Kalka C, Masuda H, Takahashi T, et al. Transplantation of ex vivo expanded endothelial progenitor cells for therapeutic neovascularization. *Proc Natl Acad Sci USA* 2000; 97: 3422-7.

- [15] Schmeisser A, Garlich CD, Zhang H, et al. Monocytes coexpress endothelial and macrophagocytic lineage markers and form cordlike structures in Matrigel under angiogenic conditions. *Cardiovasc Res* 2001; 49: 671-80.
- [16] Lenk K, Adams V, Lurz P, et al. Therapeutical potential of blood derived progenitor cells in patients with peripheral arterial occlusive disease and critical limb ischaemia. *Eur Heart J* 2005; 26: 1903-9.
- [17] Erbs S, Linke A, Adams V, et al. Transplantation of blood-derived progenitor cells after recanalization of chronic coronary artery occlusion: first randomized and placebo-controlled study. *Circ Res* 2005; 97: 756-62.
- [18] Auffray C, Fogg D, Garfa M, et al. Monitoring of blood vessels and tissues by a population of monocytes with patrolling behavior. *Science* 2007; 317: 666-70.
- [19] Giannotti G, Doerries C, Mocharla PS, et al. Impaired endothelial repair capacity of early endothelial progenitor cells in prehypertension: relation to endothelial dysfunction. *Hypertension* 2010; 55: 1389-97.
- [20] Hur J, Yoon CH, Kim HS, et al. Characterization of two types of endothelial progenitor cells and their different contributions to neovasclogenesis. *Arterioscler Thromb Vasc Biol* 2004; 24: 288- 93.
- [21] Hirschi KK, Ingram DA, Yoder MC. Assessing identity, phenotype, and fate of endothelial progenitor cells. *Arterioscler Thromb Vasc Biol* 2008; 28: 1584-95.
- [22] Awad O, Dedkov EI, Jiao C, Bloomer S, Tomanek RJ, Schatteman GC. Differential healing activities of CD34+ and CD14+ endothelial cell progenitors. *Arterioscler Thromb Vasc Biol* 2006; 26: 758-64.
- [23] Anghelina M, Krishnan P, Moldovan L, Moldovan NI. Monocytes and macrophages form branched cell columns in matrigel: implications for a role in neovascularization. *Stem Cells Dev* 2004; 13: 665-76.
- [24] Campagnolo P, Cesselli D, Al Haj Zen A, et al. Human adult vena saphena contains perivascular progenitor cells endowed with clonogenic and proangiogenic potential. *Circulation* 2010; 121: 1735-45.
- [25] Hagensen MK, Shim J, Falk E, Bentzon JF. Flanking recipient vasculature, not circulating progenitor cells, contributes to endothelium and smooth muscle in murine allograft vasculopathy. *Arterioscler Thromb Vasc Biol* 2011; 31: 808-13.
- [26] Bentzon JF, Weile C, Sondergaard CS, Hindkjaer J, Kassem M, Falk E. Smooth muscle cells in atherosclerosis originate from the local vessel wall and not circulating progenitor cells in ApoE knockout mice. *Arterioscler Thromb Vasc Biol* 2006; 26: 2696- 702.
- [27] Campagnolo P, Wong MM, Xu Q. Progenitor cells in arteriosclerosis: good or bad guys? *Antioxid Redox Signal* 2010; 15: 1013-27.
- [28] Klein D, Hohn HP, Kleff V, Tilki D, Ergun S. Vascular wall resident stem cells. *Histol Histopathol* 2010; 25: 681-9.

- [29] Hibbert B, Chen YX, O'Brien ER. c-kit-immunopositive vascular progenitor cells populate human coronary in-stent restenosis but not primary atherosclerotic lesions. *Am J Physiol Heart Circ Physiol* 2004; 287: H518-24.
- [30] Shaw JP, Basch R, Shamamian P. Hematopoietic stem cells and endothelial cell precursors express Tie-2, CD31 and CD45. *Blood Cells Mol Dis* 2004; 32: 168-75.
- [31] Purhonen S, Palm J, Rossi D, et al. Bone marrow-derived circulating endothelial precursors do not contribute to vascular endothelium and are not needed for tumor growth. *Proc Natl Acad Sci USA* 2008; 105: 6620-5.
- [32] Arras M, Ito WD, Scholz D, Winkler B, Schaper J, Schaper W. Monocyte activation in angiogenesis and collateral growth in the rabbit hindlimb. *J Clin Invest* 1998; 101: 40-50.
- [33] Nahrendorf M, Swirski FK, Aikawa E, et al. The healing myocardium sequentially mobilizes two monocyte subsets with divergent and complementary functions. *J Exp Med* 2007; 204: 3037-47.
- [34] Dewald O, Zymek P, Winkelmann K, et al. CCL2/monocyte chemoattractant protein-1 regulates inflammatory responses critical to healing myocardial infarcts. *Circ Res* 2005; 96: 881-9.
- [35] Doyle B, Sorajja P, Hynes B, et al. Progenitor cell therapy in a porcine acute myocardial infarction model induces cardiac hypertrophy, mediated by paracrine secretion of cardiogenic factors including TGF β 1. *Stem Cells Dev* 2008; 17: 941-51.
- [36] Korf-Klingebiel M, Kempf T, Sauer T, et al. Bone marrow cells are a rich source of growth factors and cytokines: implications for cell therapy trials after myocardial infarction. *Eur Heart J* 2008; 29:2851-8.
- [37] Roeske F, Stein A, Salameh A, et al. Activation of cardiomyocytes depending on their proximity to human bone marrow stem cells. *Thorac Cardiovasc Surg* 2011; 59: 78-84.
- [38] Nian M, Lee P, Khaper N, Liu P. Inflammatory cytokines and postmyocardial infarction remodeling. *Circ Res* 2004; 94: 1543-53.
- [39] Hori M, Nishida K. Oxidative stress and left ventricular remodelling after myocardial infarction. *Cardiovasc Res* 2009; 81:457-64.
- [40] Sun J, Li SH, Liu SM, et al. Improvement in cardiac function after bone marrow cell therapy is associated with an increase in myocardial inflammation. *Am J Physiol Heart Circ Physiol* 2009; 296: H43-50.
- [41] Al Haj Zen A, Oikawa A, Bazan-Peregrino M, Meloni M, Emanuelli C, Madeddu P. Inhibition of delta-like-4-mediated signaling impairs reparative angiogenesis after ischemia. *Circ Res* 2010; 107: 283-93.
- [42] Arras M, Strasser R, Mohri M, et al. Tumor necrosis factor- α is expressed by monocytes/macrophages following cardiac microembolization and is antagonized by cyclosporine. *Basic Res Cardiol* 1998; 93: 97-107.

- [43] Benjamin LE, Hemo I, Keshet E. A plasticity window for blood vessel remodelling is defined by pericyte coverage of the preformed endothelial network and is regulated by PDGF-B and VEGF. *Development* 1998; 125: 1591-8.
- [44] Detmar M, Brown LF, Schon MP, et al. Increased microvascular density and enhanced leukocyte rolling and adhesion in the skin of VEGF transgenic mice. *J Invest Dermatol* 1998; 111: 1-6.
- [45] Izquierdo E, Canete JD, Celis R, et al. Immature blood vessels in rheumatoid synovium are selectively depleted in response to anti-TNF therapy. *PLoS One* 2009; 4: e8131.
- [46] Thurston G, Rudge JS, Ioffe E, et al. Angiopoietin-1 protects the adult vasculature against plasma leakage. *Nat Med* 2000; 6: 460-3.
- [47] Yu J, deMuinck ED, Zhuang Z, et al. Endothelial nitric oxide synthase is critical for ischemic remodeling, mural cell recruitment, and blood flow reserve. *Proc Natl Acad Sci USA* 2005; 102:10999-1004.
- [48] Schachinger V, Erbs S, Elsasser A, et al. Intracoronary bone marrow-derived progenitor cells in acute myocardial infarction. *N Engl J Med* 2006; 355: 1210-21.
- [49] Moore XL, Lu J, Sun L, Zhu CJ, Tan P, Wong MC. Endothelial progenitor cells' "homing" specificity to brain tumors. *Gene Ther* 2004; 11: 811-8.
- [50] Oh HK, Ha JM, O E, et al. Tumor angiogenesis promoted by ex vivo differentiated endothelial progenitor cells is effectively inhibited by an angiogenesis inhibitor, TK1-2. *Cancer Res* 2007; 67: 4851-9.
- [51] Amariglio N, Hirshberg A, Scheithauer BW, et al. Donor-derived brain tumor following neural stem cell transplantation in an ataxia telangiectasia patient. *PLoS Med* 2009; 6: e1000029.
- [52] Ando T, Yujiri T, Mitani N, et al. Donor cell-derived acute myeloid leukemia after unrelated umbilical cord blood transplantation. *Leukemia* 2006; 20: 744-5.
- [53] Lin Q, Fu Q, Zhang Y, et al. Tumorigenesis in the infarcted rat heart is eliminated through differentiation and enrichment of the transplanted embryonic stem cells. *Eur J Heart Fail* 2010; 12: 1179-85.
- [54] Kiuru M, Boyer JL, O'Connor TP, Crystal RG. Genetic control of wayward pluripotent stem cells and their progeny after transplantation. *Cell Stem Cell* 2009; 4: 289-300.
- [55] McCarthy MJ, Loftus IM, Thompson MM, et al. Angiogenesis and the atherosclerotic carotid plaque: an association between symptomatology and plaque morphology. *J Vasc Surg* 1999; 30:261-8.
- [56] Winter PM, Morawski AM, Caruthers SD, et al. Molecular imaging of angiogenesis in early-stage atherosclerosis with alpha(v)beta3-integrin-targeted nanoparticles. *Circulation* 2003; 108: 2270-4.
- [57] Kolodgie FD, Gold HK, Burke AP, et al. Intraplaque hemorrhage and progression of coronary atheroma. *N Engl J Med* 2003; 349:2316-25.

- [58] Doebling LC, Heeger C, Aherrahou Z, et al. Myeloid CD34+CD13+ precursor cells transdifferentiate into chondrocyte-like cells in atherosclerotic intimal calcification. *Am J Pathol* 2010;177: 473-80.
- [59] Daub K, Langer H, Seizer P, et al. Platelets induce differentiation of human CD34+ progenitor cells into foam cells and endothelial cells. *FASEB J* 2006; 20: 2559-61.
- [60] Lemstrom KB, Krebs R, Nykanen AI, et al. Vascular endothelial growth factor enhances cardiac allograft arteriosclerosis. *Circulation* 2002; 105: 2524-30.
- [61] George J, Afek A, Abashidze A, et al. Transfer of endothelial progenitor and bone marrow cells influences atherosclerotic plaque size and composition in apolipoprotein E knockout mice. *Arterioscler Thromb Vasc Biol* 2005; 25: 2636-41.
- [62] Hu Y, Zhang Z, Torsney E, et al. Abundant progenitor cells in the adventitia contribute to atherosclerosis of vein grafts in ApoE-deficient mice. *J Clin Invest* 2004; 113: 1258-65.
- [63] Tepper OM, Galiano RD, Capla JM, et al. Human endothelial progenitor cells from type II diabetics exhibit impaired proliferation, adhesion, and incorporation into vascular structures. *Circulation* 2002; 106: 2781-6.
- [64] Vasa M, Fichtlscherer S, Aicher A, et al. Number and migratory activity of circulating endothelial progenitor cells inversely correlate with risk factors for coronary artery disease. *Circ Res* 2001; 89: E1-7.
- [65] Hill JM, Zalos G, Halcox JP, et al. Circulating endothelial progenitor cells, vascular function, and cardiovascular risk. *N Engl J Med* 2003; 348: 593-600.
- [66] Puska P. From Framingham to North Karelia: from descriptive epidemiology to public health action. *Prog Cardiovasc Dis* 2010; 53: 15-20.
- [67] Seeger FH, Tonn T, Krzossok N, Zeiher AM, Dimmeler S. Cell isolation procedures matter: a comparison of different isolation protocols of bone marrow mononuclear cells used for cell therapy in patients with acute myocardial infarction. *Eur Heart J* 2007; 28: 766-72.
- [68] Assmus B, Tonn T, Seeger FH, et al. Red blood cell contamination of the final cell product impairs the efficacy of autologous bone marrow mononuclear cell therapy. *J Am Coll Cardiol* 2010; 55: 1385-94.
- [69] Hansen-Smith FM, Morris LW, Greene AS, Lombard JH. Rapid microvessel rarefaction with elevated salt intake and reduced renal mass hypertension in rats. *Circ Res* 1996; 79: 324-30.
- [70] Erickson KK, Sundstrom JM, Antonetti DA. Vascular permeability in ocular disease and the role of tight junctions. *Angiogenesis* 2007; 10: 103-17.
- [71] Su RJ, Zhang XB, Li K, et al. Platelet-derived growth factor promotes ex vivo expansion of CD34+ cells from human cord blood and enhances long-term culture-initiating cells, non-obese diabetic/severe combined immunodeficient repopulating cells and formation of adherent cells. *Br J Haematol* 2002; 117: 735-46.

- [72] Iwaguro H, Yamaguchi J, Kalka C, et al. Endothelial progenitor cell vascular endothelial growth factor gene transfer for vascular regeneration. *Circulation* 2002; 105: 732-8.
- [73] Sen S, Merchan J, Dean J, et al. Autologous transplantation of endothelial progenitor cells genetically modified by adenoassociated viral vector delivering insulin-like growth factor-1 gene after myocardial infarction. *Hum Gene Ther* 2010; 21: 1327-34.
- [74] Kaur S, Kumar TR, Uruno A, Sugawara A, Jayakumar K, Kartha CC. Genetic engineering with endothelial nitric oxide synthase improves functional properties of endothelial progenitor cells from patients with coronary artery disease: an in vitro study. *Basic Res Cardiol* 2009; 104: 739-49.
- [75] Marrotte EJ, Chen DD, Hakim JS, Chen AF. Manganese superoxide dismutase expression in endothelial progenitor cells accelerates wound healing in diabetic mice. *J Clin Invest* 2010; 120: 4207-19.
- [76] Sorrentino SA, Bahlmann FH, Besler C, et al. Oxidant stress impairs in vivo reendothelialization capacity of endothelial progenitor cells from patients with type 2 diabetes mellitus: restoration by the peroxisome proliferator-activated receptor gamma agonist rosiglitazone. *Circulation* 2007; 116: 163-73.
- [77] Naldini L. Ex vivo gene transfer and correction for cell-based therapies. *Nat Rev Genet* 2011; 12: 301-15.
- [78] Hedman M, Hartikainen J, Yla-Herttuala S. Progress and prospects: hurdles to cardiovascular gene therapy clinical trials. *Gene Ther* 2011; 18: 743-9.
- [79] Sun Q, Silva EA, Wang A, et al. Sustained release of multiple growth factors from injectable polymeric system as a novel therapeutic approach towards angiogenesis. *Pharm Res* 2010; 27:264-71.
- [80] Assmus B, Urbich C, Aicher A, et al. HMG-CoA reductase inhibitors reduce senescence and increase proliferation of endothelial progenitor cells via regulation of cell cycle regulatory genes. *Circ Res* 2003; 92: 1049-55.
- [81] Fadini GP, Albiero M, Boscaro E, et al. Rosuvastatin stimulates clonogenic potential and anti-inflammatory properties of endothelial progenitor cells. *Cell Biol Int* 2010; 34: 709-15.
- [82] Spyridopoulos I, Haendeler J, Urbich C, et al. Statins enhance migratory capacity by upregulation of the telomere repeat-binding factor TRF2 in endothelial progenitor cells. *Circulation* 2004; 110:3136-42.
- [83] Emanuelli C, Zacheo A, Minasi A, et al. Adenovirus-mediated human tissue kallikrein gene delivery induces angiogenesis in normoperfused skeletal muscle. *Arterioscler Thromb Vasc Biol* 2000; 20: 2379-85.
- [84] Stone OA, Richer C, Emanuelli C, et al. Critical role of tissue kallikrein in vessel formation and maturation: implications for therapeutic revascularization. *Arterioscler Thromb Vasc Biol* 2009;29: 657-64.

- [85] Chao J, Shen B, Gao L, Xia CF, Bledsoe G, Chao L. Tissue kallikrein in cardiovascular, cerebrovascular and renal diseases and skin wound healing. *Biol Chem* 2010; 391: 345-55.
- [86] Krankel N, Katare RG, Siragusa M, et al. Role of kinin B2 receptor signaling in the recruitment of circulating progenitor cells with neovascularization potential. *Circ Res* 2008; 103: 1335-43.
- [87] Krankel N, Armstrong SP, McArdle CA, Dayan C, Madeddu P. Distinct kinin-induced functions are altered in circulating cells of young type 1 diabetic patients. *PLoS One* 2010; 5: e11146.
- [88] Spinetti G, Fortunato O, Cordella D, et al. Tissue kallikrein is essential for invasive capacity of circulating proangiogenic cells. *Circ Res* 2011; 108: 284-93.
- [89] Emanuelli C, Caporali A, Krankel N, Cristofaro B, Van Linthout S, Madeddu P. Type-2 diabetic *Lepr(db/db)* mice show a defective microvascular phenotype under basal conditions and an impaired response to angiogenesis gene therapy in the setting of limb ischemia. *Front Biosci* 2007; 12: 2003-12.
- [90] Hellberg C, Ostman A, Heldin CH. PDGF and vessel maturation. *Recent Results Cancer Res* 2010; 180: 103-14.
- [91] Levanon K, Varda-Bloom N, Greenberger S, et al. Vascular wall maturation and prolonged angiogenic effect by endothelial-specific platelet-derived growth factor expression. *Pathobiology* 2006; 73: 149-58.
- [92] Kupatt C, Hinkel R, Pfosser A, et al. Cotransfection of vascular endothelial growth factor-A and platelet-derived growth factor-B via recombinant adeno-associated virus resolves chronic ischemic malperfusion role of vessel maturation. *J Am Coll Cardiol* 2010; 56: 414-22.
- [93] Walker PA, Shah SK, Jimenez F, et al. Intravenous multipotent adult progenitor cell therapy for traumatic brain injury: preserving the blood brain barrier via an interaction with splenocytes. *Exp Neurol* 2010; 225: 341-52.
- [94] Zerneck A, Bidzhekov K, Noels H, et al. Delivery of microRNA-126 by apoptotic bodies induces CXCL12-dependent vascular protection. *Sci Signal* 2009; 2: ra81.
- [95] Zhang Y, Liu D, Chen X, et al. Secreted monocytic miR-150 enhances targeted endothelial cell migration. *Mol Cell* 2010; 39: 133-44.
- [96] Ogawa R, Tanaka C, Sato M, et al. Adipocyte-derived microvesicles contain RNA that is transported into macrophages and might be secreted into blood circulation. *Biochem Biophys Res Commun* 2010; 398: 723-9.
- [97] Aoki N, Yokoyama R, Asai N, et al. Adipocyte-derived microvesicles are associated with multiple angiogenic factors and induce angiogenesis in vivo and in vitro. *Endocrinology* 2010; 151:2567-76.
- [98] Al-Nedawi K, Meehan B, Kerbel RS, Allison AC, Rak J. Endothelial expression of autocrine VEGF upon the uptake of tumor-derived microvesicles containing oncogenic EGFR. *Proc Natl Acad Sci USA* 2009; 106: 3794-9.

- [99] Ferrari N, Glod J, Lee J, Kobiler D, Fine HA. Bone marrow-derived, endothelial progenitor-like cells as angiogenesis-selective gene-targeting vectors. *Gene Ther* 2003; 10: 647-56.
- [100] Di Santo S, Yang Z, Wyler von Ballmoos M, et al. Novel cell-free strategy for therapeutic angiogenesis: in vitro generated conditioned medium can replace progenitor cell transplantation. *PLoS One* 2009; 4: e5643.
- [101] Feig JE, Rong JX, Shamir R, et al. HDL promotes rapid atherosclerosis regression in mice and alters inflammatory properties of plaque monocyte-derived cells. *Proc Natl Acad Sci USA* 2011; 108: 7166-71.
- [102] Liu ML, Reilly MP, Casasanto P, McKenzie SE, Williams KJ. Cholesterol enrichment of human monocyte/macrophages induces surface exposure of phosphatidylserine and the release of biologically-active tissue factor-positive microvesicles. *Arterioscler Thromb Vasc Biol* 2007; 27: 430-5.
- [103] Su YR, Blakemore JL, Zhang Y, Linton MF, Fazio S. Lentiviral transduction of apoA1 into hematopoietic progenitor cells and macrophages: applications to cell therapy of atherosclerosis. *Arterioscler Thromb Vasc Biol* 2008; 28: 1439-46.
- [104] Laufs U, Werner N, Link A, et al. Physical training increases endothelial progenitor cells, inhibits neointima formation, and enhances angiogenesis. *Circulation* 2004; 109: 220-6.
- [105] Yoder MC. NO role in "EPC" function. *Blood* 2005; 105: 1846-7.
- [106] Ackah E, Yu J, Zoellner S, et al. Akt1/protein kinase B α is critical for ischemic and VEGF-mediated angiogenesis. *J Clin Invest* 2005; 115: 2119-27.
- [107] Kong D, Melo LG, Mangi AA, et al. Enhanced inhibition of neointimal hyperplasia by genetically engineered endothelial progenitor cells. *Circulation* 2004; 109: 1769-75.
- [108] Gulati R, Jevremovic D, Witt TA, et al. Modulation of the vascular response to injury by autologous blood-derived outgrowth endothelial cells. *Am J Physiol Heart Circ Physiol* 2004; 287: H512-7.
- [109] Zhang LN, Wilson DW, da Cunha V, et al. Endothelial NO synthase deficiency promotes smooth muscle progenitor cells in association with upregulation of stromal cell-derived factor-1 α in a mouse model of carotid artery ligation. *Arterioscler Thromb Vasc Biol* 2006; 26: 765-72.
- [110] Padfield GJ, Newby DE, Mills NL. Understanding the role of endothelial progenitor cells in percutaneous coronary intervention. *J Am Coll Cardiol* 2010; 55: 1553-65.
- [111] Duckers HJ, Soullie T, den Heijer P, et al. Accelerated vascular repair following percutaneous coronary intervention by capture of endothelial progenitor cells promotes regression of neointimal growth at long term follow-up: final results of the Healing II trial using an endothelial progenitor cell capturing stent (Genous R stent). *EuroIntervention* 2007; 3: 350-8.
- [112] Nakazawa G, Granada JF, Alviar CL, et al. Anti-CD34 antibodies immobilized on the surface of sirolimus-eluting stents enhance stent endothelialization. *JACC Cardiovasc Interv* 2010; 3: 68-75.

- [113] Low AF, Lee CH, Teo SG, et al. Effectiveness and safety of the genous endothelial progenitor cell-capture stent in acute ST-elevation myocardial infarction. *Am J Cardiol* 2011; 108: 202-5.
- [114] Lee YP, Tay E, Lee CH, et al. Endothelial progenitor cell capture stent implantation in patients with ST-segment elevation acute myocardial infarction: one year follow-up. *EuroIntervention* 2010; 5: 698-702.
- [115] Chong E, Poh KK, Liang S, et al. Two-year clinical registry follow-up of endothelial progenitor cell capture stent vs. sirolimus-eluting bioabsorbable polymer-coated stent vs. bare metal stents in patients undergoing primary percutaneous coronary intervention for ST elevation myocardial infarction. *J Interv Cardiol* 2010; 23: 1018.
- [116] Azzarelli S, Galassi AR, Grosso G, et al. Clinical and angiographic outcomes in elderly patients treated with endothelial progenitor cell capture coronary stents: results from a prospective single-center registry. *J Invasive Cardiol* 2010; 22: 594-8.
- [117] Beijk MA, Klomp M, Verouden NJ, et al. Genous endothelial progenitor cell capturing stent vs. the Taxus Liberte stent in patients with de novo coronary lesions with a high-risk of coronary restenosis: a randomized, single-centre, pilot study. *Eur Heart J* 2010; 31: 1055-64.
- [118] Bystron M, Cervinka P, Spacek R, et al. Randomized comparison of endothelial progenitor cells capture stent vs. cobalt-chromium stent for treatment of ST-elevation myocardial infarction. Six-month clinical, angiographic, and IVUS follow-up. *Catheter Cardiovasc Interv* 2010; 76: 627-31.
- [119] Damman P, Klomp M, Beijk MA, et al. Twelve-month outcomes after coronary stenting with the GenousTM bio-engineered R stent in diabetic patients from the e-HEALING Registry. *J Interv Cardiol* 2011; 24: 285-94.
- [120] Kaul U, Bhatia V, Ghose T, Gupta R, Kachru R, Singh G. Angiographic follow-up of genous bioengineered stent in acute myocardial infarction (GENAMI)-a pilot study. *Indian Heart J* 2008; 60: 532-5.
- [121] Wendel HP, Avci-Adali M, Ziemer G. Endothelial progenitor cell capture stents--hype or hope? *Int J Cardiol* 2010; 145: 115-7.
- [122] Granada JF, Inami S, Aboodi MS, et al. Development of a novel prohealing stent designed to deliver sirolimus from a biodegradable abluminal matrix. *Circ Cardiovasc Interv* 2010; 3: 257-66.
- [123] Ingram DA, Mead LE, Moore DB, Woodard W, Fenoglio A, Yoder MC. Vessel wall-derived endothelial cells rapidly proliferate because they contain a complete hierarchy of endothelial progenitor cells. *Blood* 2005; 105: 2783-6.
- [124] Ingram DA, Mead LE, Tanaka H, et al. Identification of a novel hierarchy of endothelial progenitor cells using human peripheral and umbilical cord blood. *Blood* 2004; 104: 2752-60.

- [125] Ingram DA, Krier TR, Mead LE, et al. Clonogenic endothelial progenitor cells are sensitive to oxidative stress. *Stem Cells* 2007; 25: 297-304.
- [126] Rauscher FM, Goldschmidt-Clermont PJ, Davis BH, et al. Aging, progenitor cell exhaustion, and atherosclerosis. *Circulation* 2003; 108: 457-63.
- [127] Dimmeler S, Aicher A, Vasa M, et al. HMG-CoA reductase inhibitors (statins) increase endothelial progenitor cells via the PI 3-kinase/Akt pathway. *J Clin Invest* 2001; 108: 391-7.
- [128] Chan AW, Bhatt DL, Chew DP, et al. Early and sustained survival benefit associated with statin therapy at the time of percutaneous coronary intervention. *Circulation* 2002; 105: 691-6.
- [129] Yokoyama T, Miyauchi K, Kurata T, Satoh H, Daida H. Inhibitory efficacy of pitavastatin on the early inflammatory response and neointimal thickening in a porcine coronary after stenting. *Atherosclerosis* 2004; 174: 253-9.
- [130] Fukuda D, Enomoto S, Shirakawa I, Nagai R, Sata M. Fluvastatin accelerates re-endothelialization impaired by local sirolimus treatment. *Eur J Pharmacol* 2009; 612: 87-92.
- [131] Indolfi C, Cioppa A, Stabile E, et al. Effects of hydroxymethylglutaryl coenzyme A reductase inhibitor simvastatin on smooth muscle cell proliferation in vitro and neointimal formation in vivo after vascular injury. *J Am Coll Cardiol* 2000; 35: 214-21.
- [132] Hibbert B, Ma X, Pourjabbar A, et al. Pre-procedural atorvastatin mobilizes endothelial progenitor cells: clues to the salutary effects of statins on healing of stented human arteries. *PLoS One* 2011; 6: e16413. Kränkel et al.
- [133] Hu Z, Zhang F, Yang Z, et al. Low-dose aspirin promotes endothelial progenitor cell migration and adhesion and prevents senescence. *Cell Biol Int* 2008; 32: 761-8.
- [134] Hambrecht R, Wolf A, Gielen S, et al. Effect of exercise on coronary endothelial function in patients with coronary artery disease. *N Engl J Med* 2000; 342: 454-60.
- [135] Linke A, Schoene N, Gielen S, et al. Endothelial dysfunction in patients with chronic heart failure: systemic effects of lower-limb exercise training. *J Am Coll Cardiol* 2001; 37: 392-7.
- [136] Vona M, Codeluppi GM, Iannino T, Ferrari E, Bogousslavsky J, von Segesser LK. Effects of different types of exercise training followed by detraining on endothelium-dependent dilation in patients with recent myocardial infarction. *Circulation* 2009; 119: 1601-8.
- [137] Schlager O, Giurgea A, Schuhfried O, et al. Exercise training increases endothelial progenitor cells and decreases asymmetric dimethylarginine in peripheral arterial disease: A randomized controlled trial. *Atherosclerosis* 2011; 217: 240-8.
- [138] Sonnenschein K, Horvath T, Mueller M, et al. Exercise training improves in vivo endothelial repair capacity of early endothelial progenitor cells in subjects with metabolic syndrome. *Eur J Cardiovasc Prev Rehabil* 2011 Feb 11. [Epub ahead of print].

- [139] Van Craenenbroeck EM, Hoymans VY, Beckers PJ, et al. Exercise training improves function of circulating angiogenic cells in patients with chronic heart failure. *Basic Res Cardiol* 2010; 105: 665-76.
- [140] Van Craenenbroeck EM, Beckers PJ, Possemiers NM, et al. Exercise acutely reverses dysfunction of circulating angiogenic cells in chronic heart failure. *Eur Heart J* 2010; 31: 1924-34.
- [141] Jenkins NT, Witkowski S, Spangenburg EE, Hagberg JM. Effects of acute and chronic endurance exercise on intracellular nitric oxide in putative endothelial progenitor cells: role of NADPH oxidase. *Am J Physiol Heart Circ Physiol* 2009; 297: H1798-805.
- [142] Kostka T, Draai J, Berthouze SE, Lacour JR, Bonnefoy M. Physical activity, aerobic capacity and selected markers of oxidative stress and the anti-oxidant defence system in healthy active elderly men. *Clin Physiol* 2000; 20: 185-90.
- [143] Hambrecht R, Adams V, Erbs S, et al. Regular physical activity improves endothelial function in patients with coronary artery disease by increasing phosphorylation of endothelial nitric oxide synthase. *Circulation* 2003; 107: 3152-8.
- [144] Gielen S, Adams V, Mobius-Winkler S, et al. Anti-inflammatory effects of exercise training in the skeletal muscle of patients with chronic heart failure. *J Am Coll Cardiol* 2003; 42: 861-8.
- [145] Ribeiro F, Alves AJ, Duarte JA, Oliveira J. Is exercise training an effective therapy targeting endothelial dysfunction and vascular wall inflammation? *Int J Cardiol* 2010; 141: 214-21.
- [146] Otto CM, Kuusisto J, Reichenbach DD, Gown AM, O'Brien KD. Characterization of the early lesion of 'degenerative' valvular aortic stenosis. Histological and immunohistochemical studies. *Circulation* 1994; 90: 844-53.
- [147] Mohler ER, 3rd, Gannon F, Reynolds C, Zimmerman R, Keane MG, Kaplan FS. Bone formation and inflammation in cardiac valves. *Circulation* 2001; 103: 1522-8.
- [148] Matsumoto Y, Adams V, Walther C, et al. Reduced number and function of endothelial progenitor cells in patients with aortic valve stenosis: a novel concept for valvular endothelial cell repair. *Eur Heart J* 2009; 30: 346-55.
- [149] Matsumoto Y, Adams V, Jacob S, Mangner N, Schuler G, Linke A. Regular exercise training prevents aortic valve disease in lowdensity lipoprotein-receptor-deficient mice. *Circulation* 2010; 121: 759-67.
- [150] Fadini GP, Albiero M, Menegazzo L, et al. Widespread increase in myeloid calcifying cells contributes to ectopic vascular calcification in type 2 diabetes. *Circ Res* 2011; 108: 1112-21.
- [151] Matsumoto Y, Adams V, Jacob S, Mangner N, Schuler G, Linke A. Regular exercise training prevents aortic valve disease in lowdensity lipoprotein-receptor-deficient mice. *Circulation* 2010; 121: 759-67.

- [152] Iyemere VP, Proudfoot D, Weissberg PL, Shanahan CM. Vascular smooth muscle cell phenotypic plasticity and the regulation of vascular calcification. *J Intern Med* 2006; 260: 192-210.
- [153] Esposito K, Marfella R, Ciotola M, et al. Effect of a mediterraneanstyle diet on endothelial dysfunction and markers of vascular inflammation in the metabolic syndrome: a randomized trial. *JAMA* 2004; 292: 1440-6.
- [154] Yeung DK, Leung SW, Xu YC, Vanhoutte PM, Man RY. Puerarin, an isoflavonoid derived from *Radix puerariae*, potentiates endothelium-independent relaxation via the cyclic AMP pathway in porcine coronary artery. *Eur J Pharmacol* 2006; 552: 105-11.
- [155] Goodfellow J, Bellamy MF, Ramsey MW, Jones CJ, Lewis MJ. Dietary supplementation with marine omega-3 fatty acids improve systemic large artery endothelial function in subjects with hypercholesterolemia. *J Am Coll Cardiol* 2000; 35: 265-70.
- [156] Lefevre J, Michaud SE, Haddad P, et al. Moderate consumption of red wine (cabernet sauvignon) improves ischemia-induced neovascularization in ApoE-deficient mice: effect on endothelial progenitor cells and nitric oxide. *FASEB J* 2007; 21: 3845-52.
- [157] Fraga CG, Litterio MC, Prince PD, Calabro V, Piotrkowski B, Galleano M. Cocoa flavanols: effects on vascular nitric oxide and blood pressure. *J Clin Biochem Nutr* 2011; 48: 63-7.
- [158] Huang PH, Chen YH, Tsai HY, et al. Intake of red wine increases the number and functional capacity of circulating endothelial progenitor cells by enhancing nitric oxide bioavailability. *Arterioscler Thromb Vasc Biol* 2010; 30: 869-77.
- [159] Huang PH, Tsai HY, Wang CH, et al. Moderate intake of red wine improves ischemia-induced neovascularization in diabetic mice-roles of endothelial progenitor cells and nitric oxide. *Atherosclerosis* 2010; 212: 426-35.
- [160] Xia L, Wang XX, Hu XS, et al. Resveratrol reduces endothelial progenitor cells senescence through augmentation of telomerase activity by Akt-dependent mechanisms. *Br J Pharmacol* 2008; 155: 387-94.
- [161] Heiss C, Jahn S, Taylor M, et al. Improvement of endothelial function with dietary flavanols is associated with mobilization of circulating angiogenic cells in patients with coronary artery disease. *J Am Coll Cardiol* 2010; 56: 218-24.
- [162] Suuronen EJ, Price J, Veinot JP, et al. Comparative effects of mesenchymal progenitor cells, endothelial progenitor cells, or their combination on myocardial infarct regeneration and cardiac function. *J Thorac Cardiovasc Surg* 2007; 134: 1249-58.
- [163] Tongers J, Roncalli JG, Losordo DW. Role of endothelial progenitor cells during ischemia-induced vasculogenesis and collateral formation. *Microvasc Res* 2010; 79: 200-6.
- [164] Assmus B, Schachinger V, Teupe C, et al. Transplantation of Progenitor Cells and Regeneration Enhancement in Acute Myocardial Infarction (TOPCARE-AMI). *Circulation* 2002; 106: 3009-17.

- [165] Wohrle J, Merkle N, Mailander V, et al. Results of intracoronary stem cell therapy after acute myocardial infarction. *Am J Cardiol* 2010; 105: 804-12.
- [166] Lunde K, Solheim S, Aakhus S, et al. Intracoronary injection of mononuclear bone marrow cells in acute myocardial infarction. *N Engl J Med* 2006; 355: 1199-209.
- [167] Tendera M, Wojakowski W, Ruzyllo W, et al. Intracoronary infusion of bone marrow-derived selected CD34+CXCR4+ cells and non-selected mononuclear cells in patients with acute STEMI and reduced left ventricular ejection fraction: results of randomized, multicentre Myocardial Regeneration by Intracoronary Infusion of Selected Population of Stem Cells in Acute Myocardial Infarction (REGENT) Trial. *Eur Heart J* 2009; 30: 1313-21.
- [168] Traverse JH, McKenna DH, Harvey K, et al. Results of a phase 1, randomized, double-blind, placebo-controlled trial of bone marrow mononuclear stem cell administration in patients following ST-elevation myocardial infarction. *Am Heart J* 2010; 160: 428-34.
- [169] Schaefer A, Zwadlo C, Fuchs M, et al. Long-term effects of intracoronary bone marrow cell transfer on diastolic function in patients after acute myocardial infarction: 5-year results from the randomized-controlled BOOST trial--an echocardiographic study. *Eur J Echocardiogr* 2010; 11: 165-71.
- [170] Meyer GP, Wollert KC, Lotz J, et al. Intracoronary bone marrow cell transfer after myocardial infarction: 5-year follow-up from the randomized-controlled BOOST trial. *Eur Heart J* 2009; 30: 297884.
- [171] Janssens S, Dubois C, Bogaert J, et al. Autologous bone marrow-derived stem-cell transfer in patients with ST-segment elevation myocardial infarction: double-blind, randomised controlled trial. *Lancet* 2006; 367: 113-21.
- [172] Govaert JA, Swijnenburg RJ, Schrepfer S, et al. Poor functional recovery after transplantation of diabetic bone marrow stem cells in ischemic myocardium. *J Heart Lung Transplant* 2009; 28: 1158-65 e1.
- [173] Pijnappels DA, Schalij MJ, van Tuyn J, et al. Progressive increase in conduction velocity across human mesenchymal stem cells is mediated by enhanced electrical coupling. *Cardiovasc Res* 2006; 72: 282-91.
- [174] Badorff C, Brandes RP, Popp R, et al. Transdifferentiation of blood-derived human adult endothelial progenitor cells into functionally active cardiomyocytes. *Circulation* 2003; 107: 102432.
- [175] Balsam LB, Wagers AJ, Christensen JL, Kofidis T, Weissman IL, Robbins RC. Haematopoietic stem cells adopt mature haematopoietic fates in ischaemic myocardium. *Nature* 2004; 428: 668-73.
- [176] Behfar A, Terzic A. Derivation of a cardiopoietic population from human mesenchymal stem cells yields cardiac progeny. *Nat Clin Pract Cardiovasc Med* 2006; 3 (Suppl 1): S78-82.
- [177] Fazel S, Cimini M, Chen L, et al. Cardioprotective c-kit+ cells are from the bone marrow and regulate the myocardial balance of angiogenic cytokines. *J Clin Invest* 2006; 116: 1865-77.

- [178] Gruh I, Beilner J, Blomer U, et al. No evidence of transdifferentiation of human endothelial progenitor cells into cardiomyocytes after coculture with neonatal rat cardiomyocytes. *Circulation* 2006; 113: 1326-34.
- [179] Kawamoto A, Tkebuchava T, Yamaguchi J, et al. Intramyocardial transplantation of autologous endothelial progenitor cells for therapeutic neovascularization of myocardial ischemia. *Circulation* 2003; 107: 461-8.
- [180] Matsuura K, Wada H, Nagai T, et al. Cardiomyocytes fuse with surrounding non-cardiomyocytes and reenter the cell cycle. *J Cell Biol* 2004; 167: 351-63.
- [181] Mummery CL, Davis RP, Krieger JE. Challenges in using stem cells for cardiac repair. *Sci Transl Med* 2010; 2: 27ps17.
- [182] Murry CE, Soonpaa MH, Reinecke H, et al. Haematopoietic stem cells do not transdifferentiate into cardiac myocytes in myocardial infarcts. *Nature* 2004; 428: 664-8.
- [183] Schuster MD, Kocher AA, Seki T, et al. Myocardial neovascularization by bone marrow angioblasts results in cardiomyocyte regeneration. *Am J Physiol Heart Circ Physiol* 2004; 287: H525-32.
- [184] Toma C, Pittenger MF, Cahill KS, Byrne BJ, Kessler PD. Human mesenchymal stem cells differentiate to a cardiomyocyte phenotype in the adult murine heart. *Circulation* 2002; 105: 93-8.
- [185] Kim SK, Pak HN, Park JH, et al. Cardiac cell therapy with mesenchymal stem cell induces cardiac nerve sprouting, angiogenesis, and reduced connexin43-positive gap junctions, but concomitant electrical pacing increases connexin43-positive gap junctions in canine heart. *Cardiol Young* 2010; 20: 308-17.
- [186] Behfar A, Yamada S, Crespo-Diaz R, et al. Guided cardiopoiesis enhances therapeutic benefit of bone marrow human mesenchymal stem cells in chronic myocardial infarction. *J Am Coll Cardiol* 2010; 56: 721-34.
- [187] Schachinger V, Assmus B, Britten MB, et al. Transplantation of progenitor cells and regeneration enhancement in acute myocardial infarction: final one-year results of the TOPCARE-AMI Trial. *J Am Coll Cardiol* 2004; 44: 1690-9.
- [188] Kawamoto A, Iwasaki H, Kusano K, et al. CD34-positive cells exhibit increased potency and safety for therapeutic neovascularization after myocardial infarction compared with total mononuclear cells. *Circulation* 2006; 114: 2163-9.
- [189] Iwasaki H, Kawamoto A, Ishikawa M, et al. Dose-dependent contribution of CD34-positive cell transplantation to concurrent vasculogenesis and cardiomyogenesis for functional regenerative recovery after myocardial infarction. *Circulation* 2006; 113: 1311-25.
- [190] Perin EC, Lopez J. Methods of stem cell delivery in cardiac diseases. *Nat Clin Pract Cardiovasc Med* 2006; 3 (Suppl 1): S1103.

- [191] Hou D, Youssef EA, Brinton TJ, et al. Radiolabeled cell distribution after intramyocardial, intracoronary, and interstitial retrograde coronary venous delivery: implications for current clinical trials. *Circulation* 2005; 112 (9 Suppl): 150-6.
- [192] Graham JJ, Foltz WD, Vaags AK, et al. Long-term tracking of bone marrow progenitor cells following intracoronary injection post-myocardial infarction in swine using MRI. *Am J Physiol Heart Circ Physiol* 2010; 299: H125-33.
- [193] Godier-Furnemont AF, Martens TP, Koeckert MS, et al. Composite scaffold provides a cell delivery platform for cardiovascular repair. *Proc Natl Acad Sci USA* 2011; 108: 7974-9.
- [194] Kaminski A, Klopsch C, Mark P, et al. Autologous valve replacement-CD133+ stem cell-plus-fibrin composite-based sprayed cell seeding for intraoperative heart valve tissue engineering. *Tissue Eng Part C Methods* 2011; 17: 299-309.
- [195] Li TS, Cheng K, Lee ST, et al. Cardiospheres recapitulate a nichelike microenvironment rich in stemness and cell-matrix interactions, rationalizing their enhanced functional potency for myocardial repair. *Stem Cells* 2010; 28: 2088-98.
- [196] Zhang Y, Thorn S, DaSilva JN, et al. Collagen-based matrices improve the delivery of transplanted circulating progenitor cells: development and demonstration by ex vivo radionuclide cell labeling and in vivo tracking with positron-emission tomography. *Circ Cardiovasc Imaging* 2008; 1: 197-204.
- [197] Penn MS, Mangi AA. Genetic enhancement of stem cell engraftment, survival, and efficacy. *Circ Res* 2008; 102: 1471-82.
- [198] Haider H, Ashraf M. Strategies to promote donor cell survival: combining preconditioning approach with stem cell transplantation. *J Mol Cell Cardiol* 2008; 45: 554-66.
- [199] Tongers J, Losordo DW, Landmesser U. Stem and progenitor cellbased therapy in ischaemic heart disease: promise, uncertainties, and challenges. *Eur Heart J* 2011; 32: 1197-206.
- [200] Sun L, Zhang T, Lan X, Du G. Effects of stem cell therapy on left ventricular remodeling after acute myocardial infarction: a metaanalysis. *Clin Cardiol* 2010; 33: 296-302.
- [201] Bai Y, Sun T, Ye P. Age, gender and diabetic status are associated with effects of bone marrow cell therapy on recovery of left ventricular function after acute myocardial infarction: a systematic review and meta-analysis. *Ageing Res Rev* 2010; 9: 418-23.
- [202] Siu CW, Liao SY, Liu Y, Lian Q, Tse HF. Stem cells for myocardial repair. *Thromb Haemost* 2010; 104: 6-12.
- [203] Roncalli J, Mouquet F, Piot C, et al. Intracoronary autologous mononucleated bone marrow cell infusion for acute myocardial infarction: results of the randomized multicenter BONAMI trial. *Eur Heart J* 2011; 32: 1748-57.

- [204] Ayach BB, Yoshimitsu M, Dawood F, et al. Stem cell factor receptor induces progenitor and natural killer cell-mediated cardiac survival and repair after myocardial infarction. *Proc Natl Acad Sci USA* 2006; 103: 2304-9.
- [205] Williams AR, Trachtenberg B, Velazquez DL, et al. Intramyocardial stem cell injection in patients with ischemic cardiomyopathy: functional recovery and reverse remodeling. *Circ Res* 2011; 108: 792-6.
- [206] Henrich D, Seebach C, Wilhelm K, Marzi I. High dosage of simvastatin reduces TNF-alpha-induced apoptosis of endothelial progenitor cells but fails to prevent apoptosis induced by IL-1beta in vitro. *J Surg Res* 2007; 142: 13-9.
- [207] Deten A, Volz HC, Briest W, Zimmer HG. Cardiac cytokine expression is upregulated in the acute phase after myocardial infarction. *Experimental studies in rats. Cardiovasc Res* 2002; 55: 329-40.
- [208] Zhang M, Methot D, Poppa V, Fujio Y, Walsh K, Murry CE. Cardiomyocyte grafting for cardiac repair: graft cell death and antideath strategies. *J Mol Cell Cardiol* 2001; 33: 907-21.
- [209] Cesari F, Sofi F, Caporale R, et al. Relationship between exercise capacity, endothelial progenitor cells and cytochemokines in patients undergoing cardiac rehabilitation. *Thromb Haemost* 2009; 101: 521-6.
- [210] Zhang Y, Ingram DA, Murphy MP, et al. Release of proinflammatory mediators and expression of proinflammatory adhesion molecules by endothelial progenitor cells. *Am J Physiol Heart Circ Physiol* 2009; 296: H1675-82.
- [211] Li TS, Takahashi M, Suzuki R, et al. Pravastatin improves remodeling and cardiac function after myocardial infarction by an antiinflammatory mechanism rather than by the induction of angiogenesis. *Ann Thorac Surg* 2006; 81: 2217-25.
- [212] Hambrecht R, Gielen S, Linke A, et al. Effects of exercise training on left ventricular function and peripheral resistance in patients with chronic heart failure: a randomized trial. *JAMA* 2000; 283: 3095-101.
- [213] Villa A, Sanchez PL, Fernandez-Aviles F. Ventricular arrhythmias following intracoronary bone marrow stem cell transplantation. *Europace* 2007; 9: 1222-3. Kränkel et al.
- [214] Price MJ, Chou CC, Frantzen M, et al. Intravenous mesenchymal stem cell therapy early after reperfused acute myocardial infarction improves left ventricular function and alters electrophysiologic properties. *Int J Cardiol* 2006; 111: 231-9.
- [215] Gepstein L. Electrophysiologic implications of myocardial stem cell therapies. *Heart Rhythm* 2008; 5(6 Suppl): S48-52.
- [216] de Boer TP, van der Heyden MA, Rook MB, et al. Proarrhythmogenic potential of immature cardiomyocytes is triggered by low coupling and cluster size. *Cardiovasc Res* 2006; 71: 704-14.

- [217] Itzhaki I, Schiller J, Beyar R, Satin J, Gepstein L. Calcium handling in embryonic stem cell-derived cardiac myocytes: of mice and men. *Ann N Y Acad Sci.* 2006 Oct; 1080: 207-15.
- [218] Schroder EA, Wei Y, Satin J. The developing cardiac myocyte: maturation of excitability and excitation-contraction coupling. *Ann N Y Acad Sci* 2006; 1080: 63-75.
- [219] Halbach M, Pfannkuche K, Pillekamp F, et al. Electrophysiological maturation and integration of murine fetal cardiomyocytes after transplantation. *Circ Res* 2007; 101: 484-92.
- [220] Zhang YM, Hartzell C, Narlow M, Dudley SC, Jr. Stem cell-derived cardiomyocytes demonstrate arrhythmic potential. *Circulation* 2002; 106: 1294-9.
- [221] Bocchi L, Savi M, Graiani G, et al. Growth factor-induced mobilization of cardiac progenitor cells reduces the risk of arrhythmias, in a rat model of chronic myocardial infarction. *PLoS One* 2011; 6: e17750.
- [222] Katritsis DG, Sotiropoulou P, Giazitzoglou E, Karvouni E, Papamichail M. Electrophysiological effects of intracoronary transplantation of autologous mesenchymal and endothelial progenitor cells. *Europace* 2007; 9: 167-71.
- [223] Gardiwal A, Reissmann LM, Kotlarz D, et al. Arrhythmia susceptibility in mice after therapy with beta-catenin-transduced hematopoietic progenitor cells after myocardial ischemia/reperfusion. *Cardiology* 2009; 114: 199-207.
- [224] Leistner DM, Schmitt J, Palm S, et al. Intracoronary administration of bone marrow-derived mononuclear cells and arrhythmic events in patients with chronic heart failure. *Eur Heart J* 2011; 32: 485-91.
- [225] Ziebart T, Yoon CH, Trepels T, et al. Sustained persistence of transplanted proangiogenic cells contributes to neovascularization and cardiac function after ischemia. *Circ Res* 2008; 103: 1327-34.
- [226] Forest VF, Tirouvanziam AM, Perigaud C, et al. Cell distribution after intracoronary bone marrow stem cell delivery in damaged and undamaged myocardium: implications for clinical trials. *Stem Cell Res Ther* 2010; 1: 4.
- [227] Tran N, Li Y, Maskali F, et al. Short-term heart retention and distribution of intramyocardial delivered mesenchymal cells within necrotic or intact myocardium. *Cell Transplant* 2006; 15: 351-8.
- [228] Goussetis E, Manginas A, Koutelou M, et al. Intracoronary infusion of CD133+ and CD133-CD34+ selected autologous bone marrow progenitor cells in patients with chronic ischemic cardiomyopathy: cell isolation, adherence to the infarcted area, and body distribution. *Stem Cells* 2006; 24: 2279-83.
- [229] Dow J, Simkhovich BZ, Kedes L, Kloner RA. Washout of transplanted cells from the heart: a potential new hurdle for cell transplantation therapy. *Cardiovasc Res* 2005; 67: 301-7.

- [230] Hayashi M, Li TS, Ito H, Mikamo A, Hamano K. Comparison of intramyocardial and intravenous routes of delivering bone marrow cells for the treatment of ischemic heart disease: an experimental study. *Cell Transplant* 2004; 13: 639-47.
- [231] Terrovitis JV, Smith RR, Marban E. Assessment and optimization of cell engraftment after transplantation into the heart. *Circ Res* 2010; 106: 479-94.
- [232] Gee AP, Richman S, Durett A, et al. Multicenter cell processing for cardiovascular regenerative medicine applications: the Cardiovascular Cell Therapy Research Network (CCTRn) experience. *Cytotherapy* 2010; 12: 684-91.
- [233] Tendera M, Wojakowski W. How to measure the effects of the intracoronary stem cell therapy? *Eur J Echocardiogr* 2010; 11: 4389.
- [234] Frederick JR, Fitzpatrick JR 3rd, McCormick RC, et al. Stromal cell-derived factor-1alpha activation of tissue-engineered endothelial progenitor cell matrix enhances ventricular function after myocardial infarction by inducing neovasculogenesis. *Circulation* 2010; 122(11 Suppl): S107-17.
- [235] Caspi O, Lesman A, Basevitch Y, et al. Tissue engineering of vascularized cardiac muscle from human embryonic stem cells. *Circ Res* 2007; 100: 263-72.
- [236] Vunjak-Novakovic G, Tandon N, Godier A, et al. Challenges in cardiac tissue engineering. *Tissue Eng Part B Rev* 2010; 16: 16987.
- [237] Kreutziger KL, Murry CE. Engineered human cardiac tissue. *Pediatr Cardiol* 2011; 32: 334-41.
- [238] Martinez EC, Kofidis T. Myocardial tissue engineering: the quest for the ideal myocardial substitute. *Expert Rev Cardiovasc Ther* 2009; 7: 921-8.
- [239] Stevens KR, Kreutziger KL, Dupras SK, et al. Physiological function and transplantation of scaffold-free and vascularized human cardiac muscle tissue. *Proc Natl Acad Sci USA* 2009; 106: 16568-73.
- [240] Zakharova L, Mastroeni D, Mutlu N, et al. Transplantation of cardiac progenitor cell sheet onto infarcted heart promotes cardiogenesis and improves function. *Cardiovasc Res* 2010; 87: 409.
- [241] Shimizu T, Sekine H, Yang J, et al. Polysurgery of cell sheet grafts overcomes diffusion limits to produce thick, vascularized myocardial tissues. *FASEB J* 2006; 20: 708-10.
- [242] Van Laake LW, Van Donselaar EG, Monshouwer-Kloots J, et al. Extracellular matrix formation after transplantation of human embryonic stem cell-derived cardiomyocytes. *Cell Mol Life Sci* 2010; 67: 277-90.
- [243] Barash Y, Dvir T, Tandeitnik P, Ruvinov E, Guterman H, Cohen S. Electric field stimulation integrated into perfusion bioreactor for cardiac tissue engineering. *Tissue Eng Part C Methods* 2010; 16: 1417-26.
- [244] Mauritz C, Schwanke K, Reppel M, et al. Generation of functional murine cardiac myocytes from induced pluripotent stem cells. *Circulation* 2008; 118: 507-17.

[245] Burridge PW, Thompson S, Millrod MA, et al. A universal system for highly efficient cardiac differentiation of human induced pluripotent stem cells that eliminates interline variability. *PLoS One* 2011; 6: e18293.

[246] Pfannkuche K, Liang H, Hannes T, et al. Cardiac myocytes derived from murine reprogrammed fibroblasts: intact hormonal regulation, cardiac ion channel expression and development of contractility. *Cell Physiol Biochem* 2009; 24: 73-86.

[247] Gore A, Li Z, Fung HL, et al. Somatic coding mutations in human induced pluripotent stem cells. *Nature* 2011; 471: 63-7.

[248] Lister R, Pelizzola M, Kida YS, et al. Hotspots of aberrant epigenomic reprogramming in human induced pluripotent stem cells. *Nature* 2011; 471: 68-73.

[249] Thum T, Hoerber S, Froese S, et al. Age-dependent impairment of endothelial progenitor cells is corrected by growth-hormonemediated increase of insulin-like growth-factor-1. *Circ Res* 2007; 100: 434-43.

[250] Fazel S, Chen L, Weisel RD, et al. Cell transplantation preserves cardiac function after infarction by infarct stabilization: augmentation by stem cell factor. *J Thorac Cardiovasc Surg* 2005; 130: 1310.

[251] Zemani F, Silvestre JS, Fauvel-Lafeve F, et al. Ex vivo priming of endothelial progenitor cells with SDF-1 before transplantation could increase their proangiogenic potential. *Arterioscler Thromb Vasc Biol* 2008; 28: 644-50.

[252] Ma FX, Chen F, Ren Q, Han ZC. Lovastatin restores the function of endothelial progenitor cells damaged by oxLDL. *Acta Pharmacol Sin* 2009; 30: 545-52.

[253] Jiang M, Wang B, Wang C, et al. Angiogenesis by transplantation of HIF-1 alpha modified "EPCs" into ischemic limbs. *J Cell Biochem* 2008; 103: 321-34.

[254] Herrler T, Leicht SF, Huber S, et al. Prostaglandin E positively modulates endothelial progenitor cell homeostasis: an advanced treatment modality for autologous cell therapy. *J Vasc Res* 2009; 46: 333-46.

[255] Chen TG, Chen JZ, Xie XD. Effects of aspirin on number, activity and inducible nitric oxide synthase of endothelial progenitor cells from peripheral blood. *Acta Pharmacol Sin* 2006; 27: 430-6.

[256] Irhimeh MR, Fitton JH, Lowenthal RM. Fucoidan ingestion increases the expression of CXCR4 on human CD34+ cells. *Exp Hematol* 2007; 35: 989-94.

[257] Zemani F, Benisvy D, Galy-Fauroux I, et al. Low-molecularweight fucoidan enhances the proangiogenic phenotype of endothelial progenitor cells. *Biochem Pharmacol* 2005; 70: 116775.

[258] George J, Goldstein E, Abashidze A, et al. Erythropoietin promotes endothelial progenitor cell proliferative and adhesive properties in a PI 3-kinase-dependent manner. *Cardiovasc Res* 2005; 68: 299-306.

- [259] Urbich C, Knau A, Fichtlscherer S, et al. FOXO-dependent expression of the proapoptotic protein Bim: pivotal role for apoptosis signaling in endothelial progenitor cells. *FASEB J* 2005; 19: 974-6.
- [260] Yamaguchi J, Kusano KF, Masuo O, et al. Stromal cell-derived factor-1 effects on ex vivo expanded endothelial progenitor cell recruitment for ischemic neovascularization. *Circulation* 2003; 107: 1322-8.
- [261] Zheng H, Shen CJ, Qiu FY, Zhao YB, Fu GS. Stromal cell-derived factor 1alpha reduces senescence of endothelial progenitor subpopulation in lectin-binding and DiLDL-uptaking cell through telomerase activation and telomere elongation. *J Cell Physiol* 2010; 223: 757-63.
- [262] Imanishi T, Hano T, Nishio I. Estrogen reduces endothelial progenitor cell senescence through augmentation of telomerase activity. *J Hypertens* 2005; 23: 1699-706.
- [263] Hibbert B, Ma X, Pourdjabbar A, et al. Inhibition of endothelial progenitor cell glycogen synthase kinase-3beta results in attenuated neointima formation and enhanced re-endothelialization after arterial injury. *Cardiovasc Res* 2009; 83: 16-23.
- [264] Mias C, Trouche E, Seguelas MH, et al. Ex vivo pretreatment with melatonin improves survival, proangiogenic/mitogenic activity, and efficiency of mesenchymal stem cells injected into ischemic kidney. *Stem Cells* 2008; 26: 1749-57.
- [265] Sasaki K, Heeschen C, Aicher A, et al. Ex vivo pretreatment of bone marrow mononuclear cells with endothelial NO synthase enhancer AVE9488 enhances their functional activity for cell therapy. *Proc Natl Acad Sci USA* 2006; 103: 14537-41.
- [266] Ceradini DJ, Yao D, Grogan RH, et al. Decreasing intracellular superoxide corrects defective ischemia-induced new vessel formation in diabetic mice. *J Biol Chem* 2008; 283: 10930-8.
- [267] Niessner A, Richter B, Penka M, et al. Endurance training reduces circulating inflammatory markers in persons at risk of coronary events: impact on plaque stabilization? *Atherosclerosis* 2006; 186: 160-5.
- [268] Steiner S, Niessner A, Ziegler S, et al. Endurance training increases the number of endothelial progenitor cells in patients with cardiovascular risk and coronary artery disease. *Atherosclerosis* 2005; 181: 305-10.
- [269] Yang YJ, Qian HY, Huang J, et al. Atorvastatin treatment improves survival and effects of implanted mesenchymal stem cells in postinfarct swine hearts. *Eur Heart J* 2008; 29: 1578-90.
- [270] Tang D, Lu J, Walterscheid JP, et al. Electronegative LDL circulating in smokers impairs endothelial progenitor cell differentiation by inhibiting Akt phosphorylation via LOX-1. *J Lipid Res* 2008; 49: 33-47.
- [271] Oikawa A, Siragusa M, Quaini F, et al. Diabetes mellitus induces bone marrow microangiopathy. *Arterioscler Thromb Vasc Biol* 2010; 30: 498-508.

[272] Shim W, Mehta A, Lim SY, et al. G-CSF for stem cell therapy in acute myocardial infarction: friend or foe? *Cardiovasc Res* 2011; 89: 20-30.

[273] Bonig H, Chudziak D, Priestley G, Papayannopoulou T. Insights into the biology of mobilized hematopoietic stem/progenitor cells through innovative treatment schedules of the CXCR4 antagonist AMD3100. *Exp Hematol* 2009; 37: 402-15 e1.

[274] Jujo K, Hamada H, Iwakura A, et al. CXCR4 blockade augments bone marrow progenitor cell recruitment to the neovasculature and reduces mortality after myocardial infarction. *Proc Natl Acad Sci USA* 2010; 107: 11008-13.

[275] Adams V, Lenk K, Linke A, et al. Increase of circulating endothelial progenitor cells in patients with coronary artery disease after exercise-induced ischemia. *Arterioscler Thromb Vasc Biol* 2004; 24: 684-90.

[276] Cubbon RM, Murgatroyd SR, Ferguson C, et al. Human exercise-induced circulating progenitor cell mobilization is nitric oxide-dependent and is blunted in South Asian men. *Arterioscler Thromb Vasc Biol* 2010; 30: 878-84.

[277] Rehman J, Li J, Parvathaneni L, et al. Exercise acutely increases circulating endothelial progenitor cells and monocyte/macrophage-derived angiogenic cells. *J Am Coll Cardiol* 2004; 43: 2314-8.

[278] Sandri M, Adams V, Gielen S, et al. Effects of exercise and ischemia on mobilization and functional activation of blood-derived progenitor cells in patients with ischemic syndromes: results of 3 randomized studies. *Circulation* 2005; 111: 3391-9.

[279] Hamed S, Alshiek J, Aharon A, Brenner B, Roguin A. Red wine consumption improves in vitro migration of endothelial progenitor cells in young, healthy individuals. *Am J Clin Nutr* 2010; 92: 1619.

[280] Urao N, Okigaki M, Yamada H, et al. Erythropoietin-mobilized endothelial progenitors enhance reendothelialization via Akt-endothelial nitric oxide synthase activation and prevent neointimal hyperplasia. *Circ Res* 2006; 98: 1405-13.

[281] Cacciatore F, Bruzzese G, Vitale DF, et al. Effects of ACE inhibition on circulating endothelial progenitor cells, vascular damage, and oxidative stress in hypertensive patients. *Eur J Clin Pharmacol* 2011; 67: 877-83.

[282] Zaruba MM, Theiss HD, Vallaster M, et al. Synergy between CD26/DPP-IV inhibition and G-CSF improves cardiac function after acute myocardial infarction. *Cell Stem Cell* 2009; 4: 313-23.

[283] Tian C, Bagley J, Forman D, Iacomini J. Inhibition of CD26 peptidase activity significantly improves engraftment of retrovirally transduced hematopoietic progenitors. *Gene Ther* 2006; 13: 652-8.

[284] Aoki J, Serruys PW, van Beusekom H, et al. Endothelial progenitor cell capture by stents coated with antibody against CD34: the HEALING-FIM (Healthy Endothelial Accelerated Lining Inhibits Neointimal Growth-First In Man) Registry. *J Am Coll Cardiol* 2005; 45: 1574-9.

[285] Garg S, Duckers HJ, Serruys PW. Endothelial progenitor cell capture stents: will this technology find its niche in contemporary practice? *Eur Heart J* 2010; 31: 1032-5.

[286] Hibbert B, Ma X, Pourdjabbar A, et al. Pre-procedural atorvastatin mobilizes endothelial progenitor cells: clues to the salutary effects of statins on healing of stented human arteries. *PLoS One* 2011; 6: e16413.

[287] Erbs S, Beck EB, Linke A, et al. High-dose rosuvastatin in chronic heart failure promotes vasculogenesis, corrects endothelial function, and improves cardiac remodeling--results from a randomized, double-blind, and placebo-controlled study. *Int J Cardiol* 2011; 146: 56-63.